



Original Research Article

DOI: 10.26479/2018.0404.43

## SYNTHESIS AND ANTIMITOTIC ACTIVITY OF CHLORO SUBSTITUTED CYANOACETYL HYDRAZONE DERIVATIVES

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**ABSTRACT:** The aim of this study were to synthesize and antimitotic activity of the cyanoacetyl hydrazones derivatives. The synthesized compounds were subjected to antimitotic studies by alliumcepa root meristamatic cells. The mitotic activity was observed in various concentrations of cyanoacetyl hydrazones. Antimitotic – activity of the synthesized compounds were observed in higher concentrations. Antimitotic activity of cyanoacetyl hydrazones were dose dependent. Increase in the concentrations is directly proportional to the activity.

**KEYWORDS:** cyanoacetyl hydrazones, meristamatic cells, MI (mitotic index), antimitotic activity, allium cepa.

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### 1. INTRODUCTION

Heterocycles form by far the largest of the classical divisions of organic chemistry. Moreover, they are of immense importance not only both biologically and industrially but also to the functioning of any developed human society as well. The majority of pharmaceutical products that mimic natural products with biological activity are heterocycles. Synthetic heterocyclic compounds can and do participate in chemical reactions in the human body. Moreover, all biological processes are expressed through chemical reaction. Such fundamental manifestation of life as the provision of energy, transmission of nerve impulses, sight, metabolisms and transfer of genetic information are all based on chemical interactions involving participation of many heterocyclic compounds such as vitamins, enzymes. The present study was designed to examine the effect of chloro substituted cyanoacetyl hydrazones on cell divisions in the root meristems of allium cepa to reveal the cytotoxic

## 2. MATERIALS AND METHODS

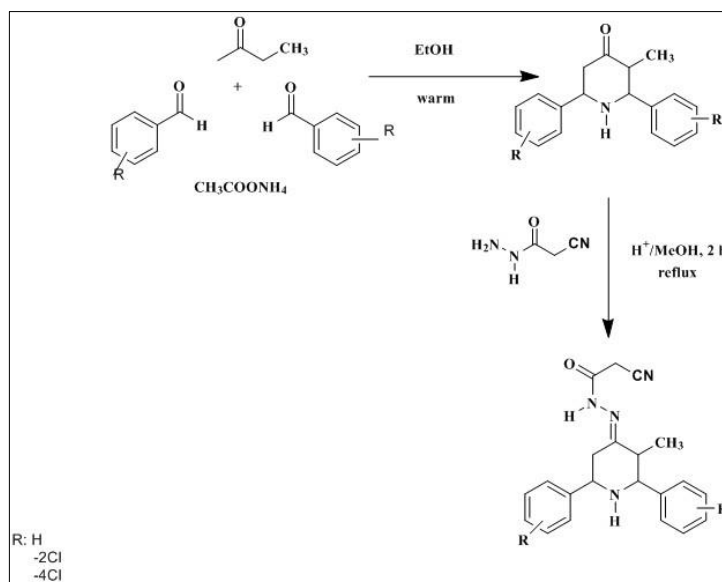
Chemicals were from E. Merck (India). IR (KBr) spectra were recorded on a Perkin-Elmer 157 infrared spectrometer ( $\nu$  in  $\text{cm}^{-1}$ ) and NMR spectra were recorded on a Bruker spectrometer DPX-300MHz (Bruker, Germany) by using  $\text{CDCl}_3$  as solvent with TMS as an internal standard.

### Preparation of S1, S2 and S3

3-methyl-2,6-diphenylpiperidin-4-one was prepared by adopting the literature method [1-6].

### Preparation of 3-methyl-2,6-diphenylpiperidin-4-one cyanoacetyl hydrazone

A mixture of 3-methyl-2,6-diphenylpiperidin-4-one (0.1 mol), cyanoacetic hydrazide (0.1 mol) in the presence of few drops of concentrated acetic acid in methanol was refluxed for 2 hours. After the completion of reaction, the reaction mixture was cooled to room temperature. The solid washed with warm water and recrystallized by methanol to afford 3-methyl-2,6-diphenylpiperidin-4-one cyanoacetyl hydrazone.



Scheme 1

### Determination of antimutagenic activity

#### Evaluation of antimutagenic activity using *Allium cepa* roots

Antimutagenic activity study was carried out by lit method [7-9].

#### Exposure to test samples

Various concentrations of S1, S2 and S3 were prepared i.e; 10  $\mu\text{g/mL}$ , 20  $\mu\text{g/mL}$ , 30  $\mu\text{g/mL}$ . The onions were divided into four groups. The first group assigned as control (tap water). Second group is allium cepa roots were dipped in S1. Third group is allium cepa roots were dipped in S2. Fourth group is allium cepa roots were dipped in S3. Fifth group is allium cepa roots were dipped in the methotrexate (0.10  $\text{mg/mL}$ ) was used as a standard control. All the groups were kept at 25°C for 96 h in direct sunlight. The root length, root number and the mitotic index were recorded after 96 h.

### Determination of MI:

Mitotic index was calculated by

$$\text{MI} = \frac{\text{Number of dividing cells}}{\text{Total number of cells}} \times 100$$

### 3. RESULTS AND DISCUSSION

S1- 3-methyl-2,6-diphenylpiperidin-4-one cyanoacetylhydrazone

S2- 3-methyl-2,6 di(bis-*o*-chloro phenyl) piperidin-4-one cyanoacetyl hydrazone

S3- 3-methyl-2,6 di(bis-*p*-chloro phenyl) piperidin-4-one cyanoacetyl hydrazone

#### Antimitotic activity of compounds (S1, S2 and S3) using *Allium cepa* root meristamatic cells

The inhibitory effect of (S1,S2 and S3) were compared with standard anticancer drug methotrexate. Increase in average mean root length (8.10mm), average mean root numbers (7) and mitotic index (87.50%) observed in control group after 96 hrs of experimental period. The average mean root length at 10, 20, and 30µg/mL of compound (S1) was 6.70mm, 3.80mm and 2.20mm at 96 hr respectively while standard shows 2.60mm. The average mean root numbers at 10, 20 and 30µg/mL of compound (S1) was 5, 4 and 3 at 96 hr respectively while standard shows 3 numbers. The mitotic index at 10, 20, and 30 µg/mL of compound (S1) was 81.55, 66.66 and 35.89% at 96 hr respectively while standard shows 36.76%. The average mean root length at 10, 20 and 30µg/mL of compound (S2) was 5.70mm, 3.70mm and 2.90mm at 96 hr respectively while standard shows 2.60mm. The average mean root numbers at 10, 20, and 30µg/mL of compound (S2) was 6, 5 and 3 at 96 hr respectively while standard shows 3 numbers. The mitotic index at 10, 20, and 30 µg/mL of compound (S2) was 75.51, 64.62 and 40.53% at 96 hr respectively while standard shows 36.76%. The average mean root length at 10, 20 and 30µg/mL of compound (S3) was 6.10mm, 4.90mm and 3.40mm at 96 hr respectively while standard shows 2.60mm. The average mean root numbers at 10, 20, and 30µg/mL of compound (S3) was 6, 4 and 4 at 96 hr respectively while standard shows 3 numbers. The mitotic index at 10, 20, and 30 µg/mL of compound (S3) was 75.82, 66.70 and 49.72% at 96 hr respectively while standard shows 36.76%. The water control shows normal growth with greater root length and numbers. Treatment with different concentrations (10, 20, and 30 µg/mL) of compounds (S1, S2 and S3) show decreased the growth gradually in dose dependent manner. The highest dose as 30µg/mL of compound (S1) has significant activity in root length, number and mitotic index and near to the standard. In the present study mitotic index of different concentrations of extract clearly indicates the efficiency in the inhibition of growth of cancer cells either by affecting

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microtubules or encouraging microtubule formation, and thus stopping the microtubules from being broken down. This makes the cells become so clogged with microtubules that they cannot continue to grow and divide [10-20]. This research has proceeded to determine the antimitotic effect of the S1, and S2 and S3. The antimitotic activity in the following order: S1> S2> S3.

**Table: 1 Effect Of Compounds (S1, S2 and S3) on MI Of *Allium cepa* roots**

Group	S1			S2			S3		
	Avg Root Growth (mm)	Avg Root Numbers	Mitotic Index (%)	Avg Root Growth (mm)	Avg Root Numbers	Mitotic Index (%)	Avg Root Growth (mm)	Avg Root Numbers	Mitotic Index (%)
Water (Control)	8.10	7	87.5	8.10	7	87.5	8.10	7	87.5
10 µg/ml	6.70	5	81.53	5.70	6	75.51	6.10	6	75.82
20 µg/ml	3.80	4	66.66	3.70	5	64.62	4.90	4	66.70
30 µg/ml	2.20	3	35.89	2.90	3	40.53	3.40	4	49.72
Std Methotrexate (0.1mg/ml)	2.60	3	36.76	2.60	3	36.76	2.60	3	36.76



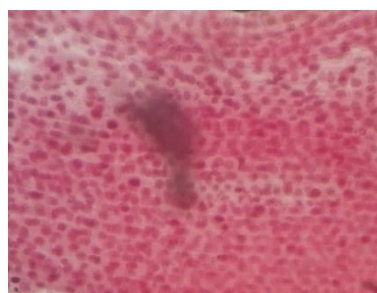
**Fig 1: Control(Water) Std (Methotrexate) (0.1mg/ml)**



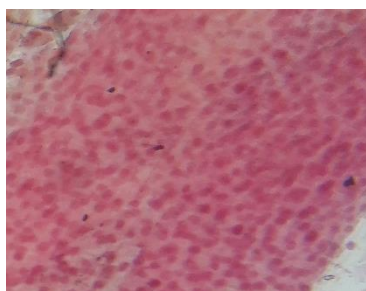
**Fig 2: 30µg/ml of compounds S1, S2, S3**



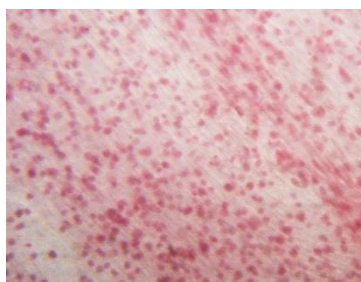
**Water(Control)**



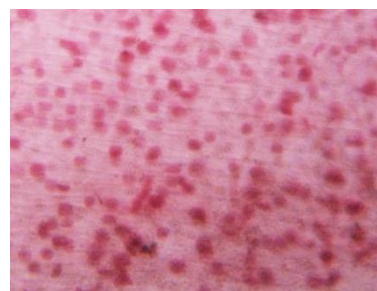
**Std (Methotrexate) (0.1mg/ml)**



**30µg/ml (S1)**



**30µg/ml (S2)**



**30µg/ml (S3)**

**Fig. 3: Photomicrograph of Compounds (S1, S2, S3) on mitotic Index of *Allium cepa***

#### 4. CONCLUSION

Synthesized a series of new chloro substituted cyanoacetyl hydrazone derivatives obtained with good yield. Among the three compounds, S1 has greater activity than S2 and S3. The antimitotic activity in the following order: S1 > S2 > S3. As a result of this cells arrest in mitosis and eventually die by apoptosis. Our findings support the reported therapeutic use of this compound as an antimitotic or anticancer agent in the Indian system of medicine.

#### ACKNOWLEDGEMENT

We are glad to express our thanks to the authorities of Harman Research Institute, Thanjavur, Tamil Nadu, India for allowing the present work to be carried out.

## CONFLICT OF INTEREST

None Declared

## REFERENCES

1. Hasan MU, Sabapathy Mohan RT and Pandiarajan K. <sup>13</sup>C and <sup>1</sup>H NMR spectral studies of some piperidin-4-one oximes. *Magnetic Resonance in Chemistry*. 1986; 24(4): 312-316.
2. Chakkaravarthy J, Muthukumar G and Pandiyarajan K. Conformational study of some N-acyl-2,6-diphenylpiperidin-4-one oximes using NMR spectra. *Journal of Molecular Structure*. 2008; 889(1): 297-307.
3. Pandiarajan K., Sekar, R, Anantharaman R, Ramalingam V. Conformational studies of some piperidin-4-ones using PMR spectroscopy. *Indian J. Chem.*1991; 30B: 490-496.
4. Tharini K and Sundaresan K. Insilico Docking studies of anti-diabetic and breast cancer activity by N'-(1-benzylpiperidin-4-ylidene)-2-cyanoacetohydrazide. *Int.J.Adv.Res.* 2017; 5(12): 601-607.
5. Tharini K and Sundaresan K. Evaluation of invitro antidiabetic activity of synthesized acetohydrazide compounds. *IOSR Journal of Applied Chemistry(IOSR-JAC)*. 2018; 11(1): 19-21.
6. Tharini K., Sundaresan K and Umamatheswari.S. Synthesis, characterization and antimutagenic activity of cyanoacetyl hydrazone derivatives. *Ejpmr*. 2017; 5(5): 314-320.
7. Fiskesjo,G.,. The Allium test-an alternative in environmental studies, the relative toxicity of metal. *Mutation Res.* 1988; 197: 243-260.
8. Shweta SS, Tapadiya GG, Lamale JJ, Khadabadi SS. Phytochemical screening and antioxidant, antimutagenic and antiproliferative activities of *Trichodesma indicum* shoot. *Ancient Sci Life*. 2014; 34: 113-121.
9. Shweta SS, Priyanka KT, Ganesh GT and Khadabadi ss. Evaluation of Phytochemical and Anticancer potential of Chloroform extract of *Trichosanthes Tricusputida* Lour Roots(Cucurbitaceae) using in-vitro models. *Int J Pharm Sci.*2013; 5(4): 203-208.
10. Grant WF. Chromosome aberration assays in *Allium*. *Mutation Res.*1982; 99: 273-291.
11. Sehgal R, Roy S and Kumar VL. Evaluation of cytotoxic potential of latex of *Calotropis procera* and podophyllotoxin in *Allium cepa* root model. *Biocell.*2006; 30(1): 9-13.
12. Badria FAR, Houssein WE, Zaghloul MG and Halim AF. Antimutagenic activity of gossypol and gossypolone. *Pharmaceutical Biol.* 2001; 39: 120 – 126.
13. Fisun. K and Rasgele. P. G. Genotoxic effects of Raxil on root tips and anthers of *Allium cepa* L. *Caryologia*. 2009; 62(1); 1-9.
14. Helsel, Z.R. Pesticide use in world agriculture. In: stout, B.A, *Energy in World agriculture*, Elsevier., New York 2, 1987: 179-195.

15. Abhang R.Y, Jogiekar P P, Kulkarni P H. Preliminary on the effect of T.cordifolia.on mitosis. Ancient sci . 1991; 1: 27.
16. Awad AB, Downie Dand Fink CS. Inhibition of growth and stimulation of apoptosis by B-sitosterol treatment of breast cancer MDA-MB-231 cells in culture. Int.J.Mol.Med. 2000; 5: 541-545.
17. Panneerselvam N, Palanikumar L and Gopinathan S. Chromosomal aberrations induced by glycidol in allium cepa L.root meristem cells. International Journal of Pharma Sciences and Research. 2012; 3(2): 300-304.
18. Kuras M, Nowakowska J, Sliwinska E and Pilarski R. Changes in chromosome structure, mitotic activity and nuclear DNA content from cell of Allium test induced by bark water extract of Uncariatementosa DC. Journal of Ethanopharmacology. 2006; 107: 211-221.
19. Williams GO and Omoh LE. Mitotic affects of aqueous leaf extract of Cymbopogo citrates in Allium cepa root tips. Cytobios; 1996: 87:161.
20. Celik TA and Aslanturk OS. Evaluation of Cytotoxcity and Genotoxicity of Inulaviscosa leaf extract with Allium teat. Journal of Biomedicine and Biotechnology. 2010; 1-8