



Original Research Article

DOI: 10.26479/2018.0406.08

MICROALGAE POPULATION IN TWO SELECTED COASTAL ZONES IN SOUTH EAST COAST OF TAMIL NADU, INDIA

Muruganantham P*, Vinoth M, Mohamed hussain J, Jeevanantham G,
Balaguru B, Khaleel Ahamed A

Department of Botany. Jamal Mohamed College (Autonomous), Tiruchirappalli, Tamil Nadu., India.

ABSTRACT: Seasonal variation and diversity of microalgae were carried out in summer, winter and monsoon seasons during the year 2014 in Rameswaram (RA) and Tuticorin (TN), South East Coast of Tamil Nadu, India. A total of 53 phytoplankton species belonging to four groups namely diatoms, dinoflagellates, green algae and blue green algae were recorded at RA and TN sites. Diatoms were found to be the dominant group (30 species) followed by blue green algae (15 species), green algae (6 species) and dinoflagellates (2 species). The density of microalgae varied from 2000 cells/L to 68,000 cells/L and the maximum density was recorded during summer and the minimum in winter season.

KEYWORDS: Diversity, Microalgae, Pollution, Marine water.

Corresponding Author: Muruganantham P*

Department of Botany. Jamal Mohamed College (Autonomous). Tiruchirappalli,
Tamil Nadu, India. Email Address: muruganbiology@gmail.com

1. INTRODUCTION

Marine phytoplanktons are free floating organisms that play an important role in food chain and contribute to the primary productivity of the ocean. Seasonal pattern in phytoplankton diversity, composition, bio volume and importantly, the magnitude of primary production [15, 26]. It is used for the variation of marine ecological problems, such as per protection of biodiversity, bionetwork characters and services. Therefore, the phytoplanktons are involving the several ecological processes with influence the species diversity [27]. The composition of phytoplankton is varied based on the environmental and hydrological conditions as well as [3, 7, 9, 11] the chemical composition of water where they exist [2,4] reported that pollution affects the distribution, standing crop and chlorophyll concentration of phytoplankton; hence the phytoplanktons are of great

importance in bio monitoring of pollution [5, 30, 31]. The plankton is an vital natural marker of the water quality. Plankton studies and observation are valuable for manage of the physico-chemical and biological environment of the water in any irrigation work [28, 29]. The objective of the present study was to compare the diversity of marine microalgae in a non-polluted site (Rameswaram) and polluted site (Tuticorin), in south east coast of Tamil Nadu.

2. MATERIALS AND METHODS

Study Area

The first study area, Rameswaram, falls within the geographical coordinate's 9° 13' 50" E and 79° 11' 55" N which is adjacent to Gulf of Manner Marine Biosphere reserve selected as control site. The second study area, Tuticorin falls within the geographical coordinate's 8° 46' 15" E and 78° 10' 7" N was selected as polluted site adjacent to the Tuticorin Harbour (Location Map).

Sample collection site

Microalgal samples were collected from Rameswaram (RA) and Tuticorin (TN) area southeast Coast of India (Figure.1). Gulf of Mannar in the southeast coast of India extends from Rameswaram Island in the north to Kanyakumari in the south. It has a chain of 21 islands stretching from Mandapam to Tuticorin to a distance of 140 km along the coast. Each one of the islands is located anywhere between 2 and 10 km from the mainland. The Gulf of Mannar was set up on 18th February 1989 jointly by the Government of India and the state of Tamil Nadu. The coastal location in Tuticorin is well known for pearl, fishery and shipbuilding. It is one of the important major Ports having a number ship movement. The movement of ships and fishing operation by mechanized boats also discharge oil effluents and petrochemical products into the sea. The Thermal power station straight dumps its ash into the sea.

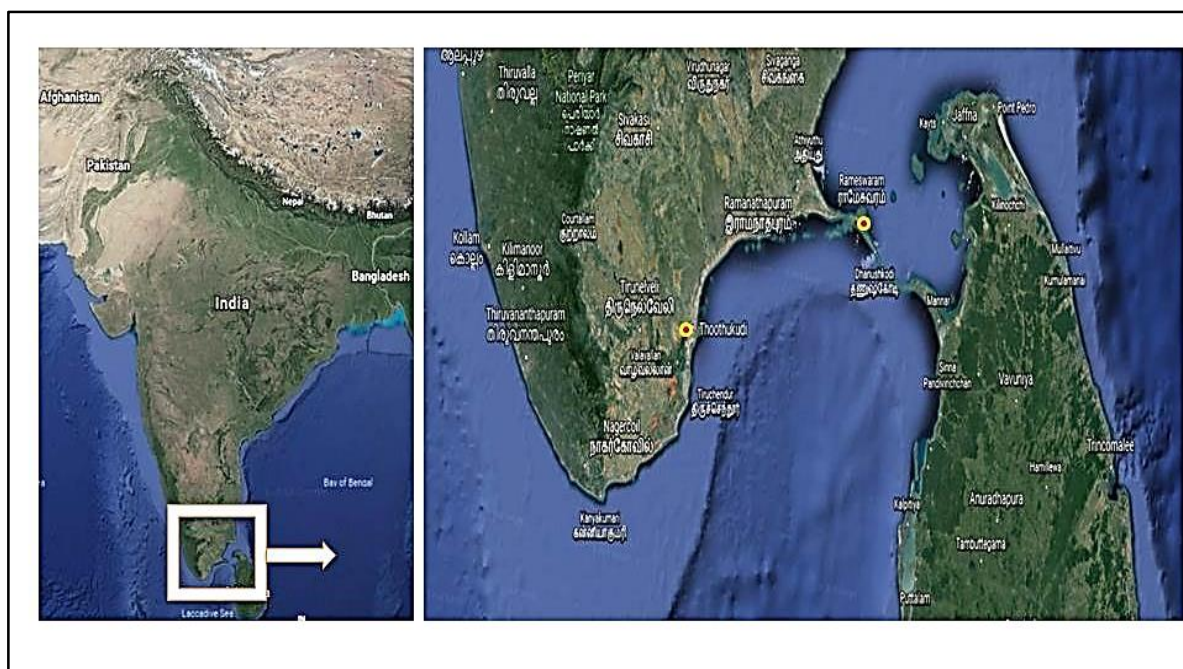


Figure 1: Sampling areas located in Rameswaram and Tuticorin of Southeast coast of Tamil Nadu, India

Sample Collection

Microalgae were collected from the surface waters in the study area during April 2014 - 2015. Samples were collected by towing plankton net (mouth diameter 45cm) made of bolting silk cloth (No.35 mesh size 30/ μm) for 20 minutes. The samples were stored in polythene bottles after preservation with Lugol's iodine solution (100:1). For the quantitative analysis of phytoplankton, the settling method as described by [38] was adopted. Numerical plankton analysis was carried out using Olympus microscope. Finally the sample was concentrated to 20 ml by decanting, and 1 drop of concentrated sample was added to Hemocytometer counting cell and were counted and identified according to standard monographs- Cyanophyta [39]. Fresh-Water Biology [40] and Indian Freshwater Microalgae [41- 49]

3. RESULTS AND DISCUSSION

This study recorded 53 species of microalgae belonging to four families' i.e. Bacillariophyceae (30), Dinophyceae (2), Cyanophyceae (15) and chlorophyceae (Table.1) *Actinoptychus splendens*, *Bacillaria paradoxa*, *Biddulphia rhombus*, *Odontella mobiliensis*, *Paralia sulcata*, *Trachynesis antillarum*, *Triceratium Robertianum*, *Oscillatoria foreau*, *Oscillatoria minnesotensis*, *Cosmarium subtumidum*, *Pediastrum tetras* and *Scenedesmus armatus* occurred in Rameswaram site. Where as *Auliscus sculptus*, *Nitzschia vitrea*, *Triceratium dubium* and *Pediastrum simplex* occurred only in TN site. *Chroococcus minutus* and *Surirella nervatus* were occurred in both the sites in all seasons. The most dominant group among microalgae was Bacillariophyceae and it contributed about 51 % of the total microalgae population. Maximum density of diatom was reported (68,000 cells L^{-1}) during summer season followed by (62,000 cells L^{-1}) monsoon season. While a minimum density with 52,000 cells L^{-1} during winter season. In TN site maximum level of density during summer season (53,000 cells L^{-1}) followed by 45,000 cells L^{-1} in monsoon season while minimum (35,000 cells L^{-1}) during winter season. Maximum density of Cyanophyceae was recorded during summer (43,000 cells L^{-1}) at RA site followed by monsoon (37,000 cells L^{-1}). While minimum cell density reported in winter (29,500 cells L^{-1}) and maximum (32,000 cells L^{-1}) during summer season followed by (26,000 cells L^{-1}) during the monsoon season. Where as in TN minimum (21,000 cells L^{-1}) during the winter season. Chlorophyceae members were the third dominant group with 7% of the total population of microalgae in the study area. In RA site maximum density was recorded during summer (15,000 cells L^{-1}) followed by monsoon (12,000 cells L^{-1}) and winter (11,500 cells L^{-1}) where as in TN site maximum density reported in (13,000 cells L^{-1}) during the monsoon followed summer (10,000 cells L^{-1}) and winter (7,000 cells L^{-1}). The fourth dominant group, Dinophyceae was represented by 3 species accounts for 4% of the total microalgae population. During summer seasons maximum density of cells was recorded (7,000 cells L^{-1}) in RA site followed by summer (6,000 cells L^{-1}) and winter. In TN site the maximum density of microalgae recorded during summer (4,000 cells L^{-1}) followed by monsoon (3,000 cells L^{-1}) and winter (2,000 cells L^{-1}).

Table 1: Number of microalgae species and the percentage composition recorded at RA and TN in study area

Group	Number of species recorded			% composition
	RA	TN	Total	
Diatoms	26	17	43	51.80723
Blue-Green algae	12	15	27	32.53012
Green algae	5	4	9	10.84337
Dinoflagellates	2	2	4	4.819277
Total	45	38	83	100

Table 2: Comparison of phytoplankton presence (✓) during different seasons at the RA and TN at Gulf of Mannar during year 2014 -2015

Species	Summer 2014		Monsoon 2014		Winter 14 – 2015	
	RA	TN	RA	TN	RA	TN
Bacillariophyceae						
<i>Actinoptychus splendens (Shadbolt)</i>	✓		✓		✓	
<i>Actinoptychus undulatus (J.W.Bailey)</i>	✓	✓		✓	✓	
<i>Amphora lineata Gregory</i>			✓			✓
<i>Auliscus caelatus Bailey</i>	✓	✓	✓	✓	✓	
<i>Auliscus sculptus (W.Smith) Brightwell</i>	✓	✓	✓	✓		✓
<i>Bacteriastrum varians Lauder</i>	✓	✓	✓	✓	✓	✓
<i>Biddulphia pulchella S.F.Gray</i>	✓	✓	✓		✓	
<i>Chaetoceros affinis Lauder</i>	✓			✓		✓
<i>Coscinodiscus asteromphalus Ehrenberg</i>	✓	✓	✓		✓	
<i>Coscinodiscus gigas Ehr</i>		✓		✓		✓
<i>Cyclotella meneghiniana Kützing</i>	✓		✓		✓	
<i>Diploneis bombus (Ehrenberg) Ehrenberg</i>	✓		✓		✓	
<i>Diploneis ovalis (Halse) Cleve</i>	✓			✓		✓
<i>Gyrosigma balticum (Ehrenberg)</i>		✓	✓	✓	✓	
<i>Mastogloia lanceolata Thwaites ex W.Smith</i>	✓				✓	
<i>Navicula granulata Ehrenberg</i>	✓		✓			

<i>Navicula lyra</i> Ehrenberg	✓	✓		✓	✓	
<i>Nitzschia plana</i> W.Smith			✓		✓	✓
<i>Paralia sulcata</i> (Ehrenberg) Cleve	✓	✓	✓	✓	✓	
<i>Pinnularia acrosphaeria</i> W.Smith	✓	✓	✓	✓		✓
<i>Pleurosigma angulatum</i> (Queckett) W.Smith	✓	✓	✓	✓	✓	✓
<i>Rhabdonema minutum</i> Kutzing	✓	✓	✓		✓	
<i>Rhizosolenia styliiformis</i> T.Brightwell	✓			✓		✓
<i>Surirella fastuosa</i> (Ehrenberg)	✓	✓	✓		✓	
<i>Surirella nervatus</i> (Grunw)		✓		✓		✓
<i>Thalassionema nitzschioides</i> (Grunow)	✓		✓		✓	
<i>Thalassiosira excentrica</i> (Ehrenberg) Cleve	✓		✓		✓	
<i>Triceratium dubium</i> Brightwell	✓	✓		✓		✓
<i>Triceratium favus</i> Ehrenberg		✓		✓		
<i>Triceratium Robertianum</i> Grey	✓				✓	
Cyanophyceae						
<i>Chroococcus</i> sp.	✓	✓		✓	✓	
<i>Gloeocapsa polydermatica</i>			✓		✓	✓
<i>Lyngbya burgertii</i>	✓	✓	✓	✓	✓	
<i>Lyngbya</i> sp.	✓	✓	✓	✓		✓
<i>Microcoleus acculismus</i>	✓	✓	✓	✓	✓	✓
<i>Phormidium foveolarum</i>	✓	✓	✓		✓	
<i>Phormidium fragile</i>	✓			✓		✓
<i>Phormidium tenue</i>	✓	✓	✓		✓	
<i>Phormidium valderianum</i>		✓		✓		✓
<i>Phormidium</i> sp.	✓		✓		✓	
<i>Phormidium mole</i>	✓		✓		✓	
<i>Phormidium uncinatum</i>	✓	✓		✓		✓
<i>Synechococcus elongatus</i>		✓	✓	✓	✓	
<i>Spirulina subsalsa</i>		✓		✓		✓
<i>Synechocystis aquatilis</i>	✓	✓	✓		✓	
Dinophyceae						
<i>Ceratium lineatum</i>			✓		✓	✓
<i>Ceratium massiliense</i>	✓	✓		✓	✓	✓

Chlorophyceae						
<i>Cosmarium subtumidium</i>	✓	✓	✓	✓	✓	✓
<i>Pediastrum boryanum</i>	✓		✓		✓	✓
<i>Pediastrum simplex</i>	✓			✓		✓
<i>Pediastrum tetras</i>	✓	✓	✓		✓	
<i>Scenedesmus armatus</i>		✓		✓		✓
<i>Scenedesmus bijugatus</i>	✓		✓		✓	

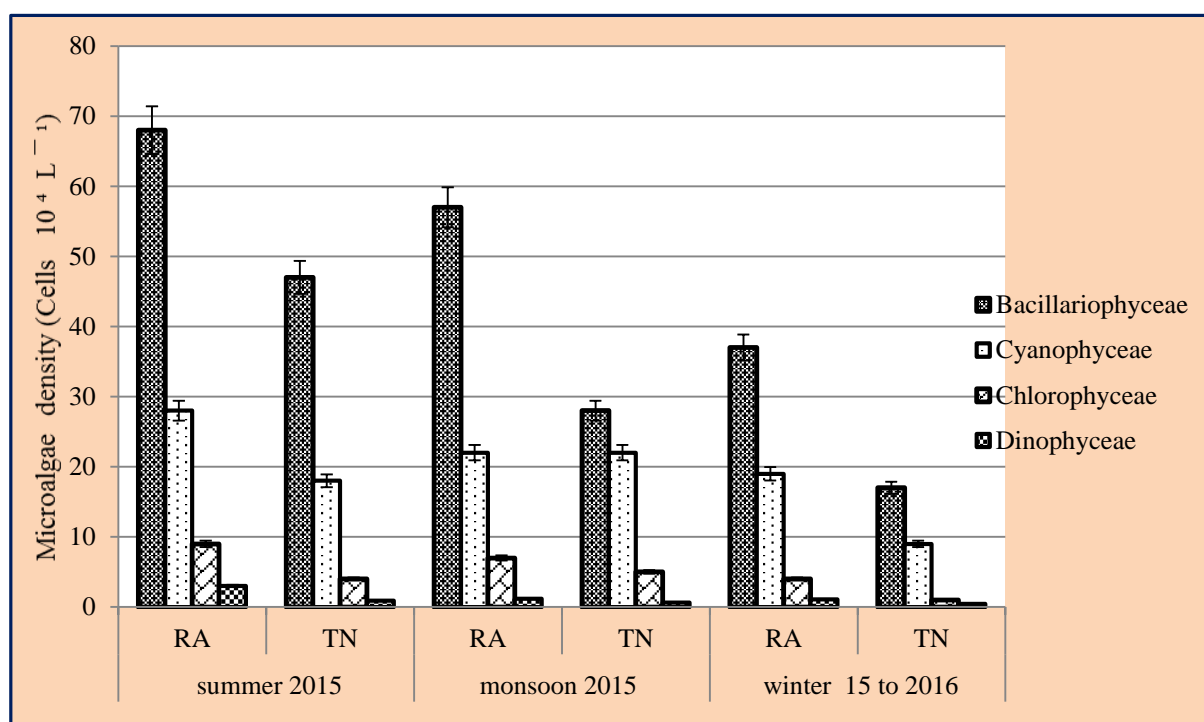


Figure 2: Seasonal variation of microalgae density observed at RA and TN during year 2014 to 2015

DISCUSSION

Diatoms are ecologically important and they are used for monitoring environmental conditions of waters. Diatoms are the dominant group in the present study and it is supported by the work of [13, 14, 20, and 21]. Diatoms are followed by dinoflagellates and blue green algae as dominant group. Similar observation were made [32] from the Palk Bay [33] from the Gopalpur [34, 35] from the Pitchavaram mangroves, [36] from the Coromandel Coast. [37] From the Pichavaram mangrove. Dominance of dinoflagellates and cyanophytes in Central Arabian Sea and Bay of Bengal [6, 8, 22]. *Amphora lineata* were recorded from the RA and TN site only during winter season. [10] Observed that *Amphora sp.* distribution during winter in Belgian coastal zone. *Cyclotella meneghiniana* was recorded only in RA site during all seasons but absent in TN site, which evident that the *C.*

meneghiniana and *Scenedesmus bijugatus* are the low pollution tolerant species replaced by *Pleurosigma angulatum*, *Bacteriastrum varians*, *Microcoleus acculismus* and *Cosmarium subtumidium*, *Actinoptychus splendens*, *Diploneis bombus*, *Mastogloia lanceolata*, *Navicula granulata*, *Coscinodiscus gigas*, *Thalassionema nitzschioides* and *Triceratium Robertianum* were reported only from RA site but their absence reported in the TN site which mainly affected by thermal power stations and other industrial pollution. This evident that diatoms are the most important primary producers of both marine and freshwater environments and regulates the ocean's silicon cycle is considerable [19, 25]. Diatoms have also been used as valuable indicators in historical assessments of water quality [16, 17, and 23]. *Triceratium dubium* is indicator for coastal wetlands [18, 24]. *T. dubium* was recorded during summer in RA and TN site, but their presence recorded during winter and monsoon seasons. The species is tolerant to thermal pollution, during monsoon and winter season the flow of water reduce the thermal pollution in RA site, hence their absence in RA site. But the TN site polluted by the thermal power plants, hence their presence indicate that the area affected by the thermal pollution.

4. CONCLUSION

From the present study the highest diversity of phytoplankton communities were observed at the RA site. The species richness increased during summer season due to positive environmental conditions and presence of nutrient abundance, but in monsoon and winter season where inflow of water is less compared with other seasons. Water quality parameter would affect the microalgae density which increased during monsoon and lesser during winter. The microalgae concentration were less during monsoon season due to the dilution factors and sudden changes of water quality parameters, which is lead to the lesser amount of photosynthetic activity by primary producers. The study reported that the microalgae played as the bio indicator of water pollution. It is concluded that TN site (Tuticorin) is most affected by pollution than the Rameswaram site (RA). Hence the comparison of the physiochemical parameters with the seasonal distribution of microalgae diversity and distribution are essential and is recommended.

ACKNOWLEDGEMENT

Author P. Muruganantham is thankful to University Grant Commission, Government of India for awarding the Project fellowship and the Principal, and the Secretary and Correspondent, Jamal Mohamed College, Tiruchirappalli for their constant encouragement and providing necessary facilities.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

REFERENCES

1. Adebowale FO, Agunbiade B and Olu-Owolabi I. Impacts of natural and anthropogenic multiple sources of pollution on the environmental conditions of Ondo State coastal waters, Nigeria, *Electronic Journal of Environment, Agriculture & Food Chemistry*. 2008.
2. Babu A, Varadharajan D, Vengadesh Perumal N, Thilagavathi B, Manikandarajan T. Diversity of Phytoplankton in Different Stations from Muthupettai, South East Coast of India. *J Marine Sci Res Dev*.2013; 3:128.
3. Barnes RSK. *Coastal Lagoons*. 2nd Edn., Cambridge University Press, London, U.K, 1980; pp: 106.
4. Davies OA, Abowei JFN and Tawari CC. Phytoplankton community of Elechi Creek, Niger Delta, Nigeria: A nutrient polluted tropical creek. *Am. J. Appl. Sci*. 2009; 6(6): 1143-1152.
5. Geetha VM and Kondalarao B. Distribution of phytoplankton in the coastal waters of East coast of India. *Indian Journal of Marine Sciences*.2004;33(3): 262-268.
6. Hulyal SB, Kaliwal BB. Dynamics of phytoplankton in relation to physico-chemical factors of Almatti reservoir of Bijapur district, Karnataka State. *Environ. Monit. Assess*. 2009; 153, 45–59.
7. Krey J. Primary production in the Indian Ocean I. in: *The Biology of Indian Ocean*, edited by B.Zeitzschel (Springer Verlag, New York). 1973; pp 115-126.
8. Lee RE. *Psychology*. Cambridge University Press, New York.1999. 614. pp.
9. Millman MC Cherrier & Ramstack J. The seasonal succession of the phytoplankton community in Ada Hayden lake, North Basin, Ames., Iowa, Limnology Laboratory, Iowa. State University, Ames, Iowa.2005.
10. Richardson K. Harmful or exceptional phytoplankton blooms in the marine ecosystem. *Adv.Mar. Biol*. 1997; 31:302-401.
11. Sarojini Y & Sarma NS. Vertical distribution of phytoplankton around Andaman & Nicobar Islands. Bay of Bengal. India. *J. Mar. Sci*. 2001.30; 65-69.
12. Sawant S, Madhupratap M. Seasonality and composition of phytoplankton in the Arabian Sea. *Curr. Sci*.1996; v.71, p.869-873.
13. Schluter M, Kraberg A, Wiltshire KH. Long-term changes in the seasonality of selected diatoms related to grazers and environmental conditions. *J Sea Res*.2012; 67: 91-97.
14. Trobajo R, Sullivan M. Applied diatom studies in estuaries and shallow coastal environments. *The Diatoms Applications for the Environmental and Earth Sciences*, Cambridge University Press, Cambridge, UK.2010; pp: 309-323.
15. Xavier Benito Granell. Benthic diatoms and foraminifera as indicators of coastal wetland habitats: application to Palaeoenvironmental reconstruction in a Mediterranean delta, Ph.D thesis. Centre for Climate Change, Geography Department, University Rovira and Virgili. 2015.

16. Yool A, Tyrrell T. Role of diatoms in regulating the ocean's silicon cycle. *Glob Biogeochem Cycles*. 2003; 17: 1103.
17. Muruganantham P, Gopalakrishnan T, Chandrasekaran R, Jeyachandran S. Seasonal Variations and Diversity of Planktonic Diatoms of Kodikkarai and Velanganni, Southeast Coast of India. *J. Ocean. And Mar. Env. Sys*. 2012; 2(1):1-0.
18. Murugaraj GN, Jeyachandran S. Effect of salinity stress on the marine diatom *Amphora coffeaeformis* (Ag.) Kuetz. *Bacillariophyceae*) in relation to proline accumulation. *Seaweed Res. Utiln*. 2007; 29:227-31.
19. Jeyachandran S. Studies on the intertidal diatoms from the Pitchavaram mangroves (India). Ph.D, Thesis Annamalai University. 1989.
20. Muruganantham P, Gopalakrishnan T, Chandrasekaran R, Jeyachandran S. Seasonal variations and diversity of planktonic diatoms of Muthupet and Aarukattuthurai, Southeast Coast of India. *Advances in Applied Science and Research*. 2012; 3(2):919-29.
21. Muruganantham P, Mohammed Hussain J, Jeevanantham G, Vinoth M, Balaguru B, Khaleel Ahamed A. Effect of physiochemical factor induced Reactive Oxygen Species (ROS) on Marine Microalgal population in South East Coast of India - A Spatial Temporal approach Haya: The Saudi Journal of Life Sciences. 2018;3(3):226-242.
22. Vinoth M, Muruganantham P, Jeevanantham G, Hussain JM, Balaguru B, Ahamed AK. Distribution of cyanobacteria in biological soil crusts in sacred groves forest of ariyalur and pudukottai districts, tamilnadu, india. *Rjlbpcs*. 2017;3(3) :215-241
23. Varadharajan D, Soundarapandian P. Biodiversity and abundance of phytoplankton from Muthupettai mangrove region, south east coast of India. *Journal of Aquaculture Research and Development*. 2015; 6(12).
24. Roelke D, Buyukates Y. Dynamics of phytoplankton succession coupled to species diversity as a system-level tool for study of *Microcystis* population dynamics in eutrophic lakes. *Limnology and Oceanography*. 2002 Jul 1; 47(4):1109-18.
25. Peerapornpisal Y, Sonthichai W, Somdee T, Mulsin P, Rott E. Water quality and phytoplankton in the Mae Kuang Udomtara Reservoir, Chiang Mai, Thailand. *J. Sci. Fac. Cmu*. 1999; 26(1):25-43.
26. Elliott JA, Irish AE, Reynolds CS. Predicting the spatial dominance of phytoplankton in a light limited and incompletely mixed eutrophic water column using the PROTECH model. *Freshwater Biology*. 2002 Mar; 47(3):433-40.
27. Diehl S. Paradoxes of enrichment: Effects of increased light versus nutrient supply on pelagic producer-grazer systems. *Am Nat*. 2007;169: E173- E191.
28. Nowrouzi S, Valavi H. Effects of environmental factors on phytoplankton abundance and diversity in Kaftar Lake. *J Fish Aquat Sci*. 2011; 6: 130-140.

29. Raghuprasad R. Plankton calendars of the inshore waters at Mandapam, with a note on the productivity of the area. *Indian J Fish.* 1958;5: 170-188.
30. Rajashree G, Panigrahy RC. Ecology of phytoplankton in coastal waters off Gopalpur, east coast of India. *Ind J Mar Sci.* 1996; 25: 13-18.
31. Mani P. Phytoplankton in pitcharavarm mangroves, eastcoast of India. *Indian J Mar Sci.* 1994; 23: 22-26.
32. Vasantha K. Studies on hydrobiology and decomposition of macrophytes in Porto Novo marine environment (Southeast coast of India), Ph.D., Thesis, Annamalai University, India. 1989; 252.
33. Govindasamy C. Coromandel Coast of India – A study on hydrobiology and heavy metals, Ph.D., Thesis, Annamalai University, India. 1992; 119.
34. Rajkumar M, Perumal P, Ashok Prabu V, Vengadesh Perumal N, Thillai Rajasekar K. Phytoplankton diversity in Pichavaram mangrove waters from south-east coast of India. *Journal Of Environmental Biology.* 2009; 30: 489-498.
35. Sukhanova ZN. Setting without the inverted microscope. In: *Phytoplankton manual*, UNESCO, (Ed: A. Sourina). Page Brothers (Nourish) Ltd. 1978; pp: 97.
36. Desikachary TV. *Cyanophyta*. New Delhi: Indian Council of Agricultural Research; 1959.
37. Edmondson WT, editor. *Ward & Whipple's Fresh-Water Biology*. Wiley; 1959.
38. Anand N. *Indian freshwater microalgae*. Bishen Singh Mahendra Pal Singh; 1998.
39. Cleve-Euler A. Die diatomeen von Schweden und Finnland. *K.Svenska vetensk. Akad. Handl., Ser.* 1951-55. pp: 4.
40. Desikachary TV. Marine fossil diatoms from India and Indian Ocean region, In: *Atlas of Diatoms* (Ed.T.V. Desikachary), Madras science foundation, Madras. 1986; pp: 77.
41. Desikachary TV. *Atlas of diatoms fascicle V (Marine diatoms of the Indian Ocean region)*, With (Marine diatoms of the Indian Ocean region). 1988; pp: 1-13
42. Subrahmanyam R. The diatoms of the Madras Coast, *Proc. Indian Acad. Sci.* 1946; 24(13): 85-197.
43. Cupp EE. Marine plankton diatoms of the west coast of North America, *Bull. Scripps Inst. Oceanogr.* 1943; 5(1): 1-237.
44. Hustedt F. Die Kieselagen Deutschlands. Osterreichs und Schweiz. In: *Rabenhorst's Kryptogam Flora*. 1927-1966; 7(1).
45. Boyer CS. Synopsis of North American Diatomaceae, *Proc Acad. Natl.Sci.phila.* 1926-27; 78(Suppl. Pt. 1): 1-228, 79(Suppl. 2): 229-583.
46. Hendey NI. *Bacillariophyceae (Diatoms)*, in an introductory account of the smaller algae of British Coastal waters, *Fishery inves. Ser., IV*, London, 1964; pp.: 317.