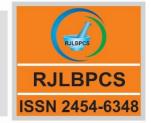


Life Science Informatics Publications

Research Journal of Life Sciences, Bioinformatics, Pharmaceutical and Chemical Sciences

Journal Home page http://www.rjlbpcs.com/



Original Review Article

DOI: 10.26479/2019.0501.31

DUST POLLUTION AND ITS INFLUENCE ON VEGETATION – A CRITICAL ANALYSIS

S. Kameswaran¹, Y. Gunavathi², P. Gopi Krishna^{2*}

1. Department of Botany, Vikrama Simhapuri University PG Centre, Kavali, AP, India.

2. Department of Zoology, Vikrama Simhapuri University PG Centre, Kavali, AP, India

Abstract: The major sources of dust pollution include suspension of soil, agriculture-related activities, road dust, vehicular exhaust, power plants, construction activities, open fires, brick kilns, cement factories and volcanoes. Due to this pollution, plants suffer from stomatal closure leading to cell/tissue changes, leaves' necrosis, pigment loses and chlorosis. The first physiological reaction after dust deposition to the vegetation takes place on the leaf with reduced net assimilation efficiency. Moreover, the long-term depositions change the photochemistry leading to retarded leaf growth. Deposits for many years over plants' surface lead to a large-scale reduction in the assimilate balance. Additionally, there are few reports on abrasive effects of dust, especially under high wind speed; supporting secondary effects such as an increase in diseases and pest incidence after the protective leaf cuticle were removed physically. Changes in soil chemistry due to dust deposition in the rhizosphere also lead to a change in soil nutritional values. In this article, we summarized the influences of dust pollution on various parameters of vegetation.

Keywords: Dusts; Air pollution; Photosynthesis; Stomatal closure; Fly ash.

Corresponding Author: Dr. P. Gopi Krishna* Ph.D.

Department of Zoology, Vikrama Simhapuri University Post-Graduate Centre, Kavali-524201, Andhra Pradesh, India. Email Address: pitchika.gopikrishna@gmail.com

1. INTRODUCTION

Most developing nations have been influenced by atmospheric pollution, in terms of human health [1], climate change [2] and loss of biodiversity [3]. With urban transport development, trafficderived pollutants become an increasing problem [4], [5], [6] and have been linked to respiratory and cardiovascular disease, birth and developmental defects, cancer and so on. According to WHO

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications [7], the life expectancy of urban residents in strongly polluted areas could decrease by over one year, particularly for children and people with lung and heart disease. There are 800000 deaths annually, which could be due to urban air pollutants [8]. The major sources of dust pollution include suspension of soil, agriculture related activities, road dust, vehicular exhaust, power plants, construction activities, brick kilns and cement factories etc. [9], [10]. In addition, transboundary and long range transport also contributes a significant amount of atmospheric dust in the south Asian region [11]. Particulate matter pollutions usually arise from multi-various sources. They can derive from anthropogenic activity as well as from natural sources [12]. They can be released into the environment as primary or secondary particles. Primary particulate matters develop and are produced directly from the sources. While secondary particulate matters are reaction products in the atmosphere. Examples of such reaction products are ammonium sulfates and nitrate of ammonia as well as aldehydes and ketones [13]. These substances adhere to other atmospheric particles easily leading to the formation of nuclei of condensation [14]. Countries like India have high loading of soil derived dust in the atmosphere under prevailing dry weather conditions. The soil derived particulate matter is rich in CaCO₃ and acts as an effective scavenger of atmospheric SO₂ forming CaSO₄ which is also removed through dust-fall [15], [16], [17]. Deposition of such particulate matter on the foliar may lead to different phytotoxic effects depending on the characteristics of the deposited material. The sulphate, nitrate and heavy metals are the most commonly reported air pollutants responsible for phytotoxic effects [18]. Sulphate and nitrate being acidic species have a higher aqueous affinity which allows these ions to mobilize into the foliar mesophyll cells creating stress [19], [20]. Dust-fall having pH values ≥ 9 might cause direct or indirect injury to the foliar tissues [21], [22]. Deposition of dust on the foliar surface may alter its optical properties, particularly the foliar surface reflectance in the visible and near infrared region [23], [24]. Dust-fall also alters optical properties of snow-cover which can lead to an increase in temperature of vegetation surface [27], [28], changes in grazing patterns of animal [25], [26], [27]. According to Sharifi et al., [28], the deposition of road dust of 40 g m⁻² can cause the 2-3°C increase in leaf temperature in desert environments. The species having stomata in grooves might be less affected than the species in which the stomata are located at the outer surface of the leaf. Hence, dust-fall on foliar surfaces has significant impacts on the photosynthesis and growth [28], [29]. Trees and shrubs are considered as efficient filters for road dust [30], [31]. In the Indian region, dust is abundant in the atmosphere. Many industrial processes, especially in quarrying causes fine particle matters producing in huge quantities around the world [32]. Other major sources of particle matters are traffic and thermal power plants [33], [34]. The contribution of different sources towards total dust pollution varies from location to location. Even within traffic, the emissions from car vary a lot [35]. Langner [35] used for his calculations of particulate matters filter efficacy rates of urban greens a mean particle matter

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications emission rate per car of 100 mg km⁻¹. It is not easy to transfer empirically based deposition factors to complex urban situations. Some silica particle matters derived from the rock in inland settings and from shells or algae in coastal areas are relatively inert in nature [36], [37], [38]. Other particulate matters like limestone quarry dust are highly alkaline in nature [39], [40]. Airborne particulates, used as an indicator of heavy metal pollution due to atmospheric deposition, mainly originates from fuel combustion by gasoline/diesel powered vehicles and non-combustion sources including vehicle brake and tire abrasion, gardening, household waste discharge, architectural painting and building structure erosion [41], [42], [43], [44] and it contains a mixture of heavy metals, black carbon, polycyclic aromatic hydrocarbons and other substances suspended in the atmosphere [45]. The roadside soil in Beijing is significantly contaminated by heavy metals [46], of which some are carcinogenic, mutagenic, teratogenic, endocrine disruptors, whereas others cause neurological and behavioral changes, especially in children [47]. Vegetation has been used as an indicator of air pollution [48], [49], as their leaves can effectively adsorb air particulates [50], [51], [52], [53], [54] and reduce air pollutants [55]. Dust can impact plants directly by covering aboveground parts of the vegetation or indirectly over the soil and the root systems. Apart from the pure size of the dust particle the physical and chemical characteristics of the particles are also important for their effect on plants. Plants differ in the ability to collect particulate matter from the air [56], [57] and in their reaction to particulate matters depositions.

Dust deposition on Plants

Generally, dust particles are filtered by plants at a much higher rate than fine particles [35]. The amount of coal dust deposited on plant surfaces varies significantly spatiotemporally. Many of the factors responsible for the coal dust deposition are similar to those governing deposition of other pollutants. Generally greater surface roughness increases deposition rate [58]. This parameter is especially important at greater wind speeds [59]. Chamberlain [59] also describes that wet surfaces can result in higher deposition rates. The plant reaction differs depending on the size of the deposited particles served that chemical interactions of dust with the vegetation surface are impaired by the presence of water. For example, fly ash to a great extent is not water soluble and thus the probability of chemical burn only very small. This may not apply to other types of dust. Pilot experiments on particulate matters which affect plants were conducted by Dugger and Cooley [60] on commercial crops. They compared charcoal, calcium carbonate, and aluminum hydroxide dusts on Lycopersicon esculentum. All three dusts increased transpiration, but charcoal reduced growth parameters while the other dusts increased it. Sree Rangaswami and Jambulingam [61] studied in southern India the distribution of plant species around a cement factory that emits dusts. Out of 54 species, they found only nine species were able to grow close to the factory. All of those plant species possessed small leaves, resulting in a reduced dust load. Generally, unpaved roads produce higher dust levels than paved roads [62]. Brown and Berg [40] undertook a detailed

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications study of an unpaved road in Alaska. He found that in the summer about 10 g m⁻² day⁻¹ was a logarithmic decline in deposition away from the road, with deposition still occurring 1 km away. Dusts affect plant physiology at both physically as well as at the chemical and biochemical level. The absolute level of dust deposition might be important for physical effects. Fine dust particles can clog stomatal openings [63], [64], reduce photosynthesis [65], increases leaf temperature [66], [67] and increase transpiration [68].

Dust Effect on Leaf Morphology

Leaf is the most sensitive part to be affected by air pollutants instead of all other plant parts such as stem and roots. The sensitivity rests on the fact that the major portions of the important physiological processes are concerned with leaf. Therefore, the leaf at its various stages of development serves as a good indicator of air pollution. Pollutants came from the auto emission can directly affect the plant by entering into the leaf, destroying individual cells, and reducing the plant ability to produce food. Urban air pollution is a major environmental problem, mainly in the developing countries [69]. The response of the plant to dust accumulation may vary according to different species, as dust deposition fluctuates with plant species due to leaf orientation, leaf surface geometry, phyllotaxy, epidermal and cuticular features, leaf pubescence, height and canopy of roadside plants [30], [70], [71], [72]. With the accumulation of dust, the roadside plant may exhibit adaptive response by changing morphological and physiological attributes. Heavy metals released from automobiles are extremely toxic and reduces plant growth and morphological parameters. Therefore, a study conducted by Ahmad et al., [73] is agreement that the Cadmium had toxicity at 5 mg L⁻¹ in case of root and shoot growth. Air pollution due to vehicular emission mostly arises from cars, buses, mini-buses, wagons, rickshaws, motorcycles and trucks. These resources introduce varieties of pollutants (oxides of nitrogen, sulphur, hydrocarbon, ozone, particulate matters, hydrogen fluoride, peroxyacyl nitrates, etc.) into the environment which not only put an adverse effect on the health of human beings, and animals, but seriously treating the trees and crops of such areas. Research studies revealed that plants growing in the urban areas are affected greatly by these pollutants [74]. Pollutants can cause leaf injury, stomatal damage, premature senescence, decrease photosynthetic activity, disturb membrane permeability and reduce growth and yield in sensitive plant species [75]. Reductions in leaf area and leaf number may be due to decreased leaf production rate and enhanced senescence. The reduced leaf area results in reduced absorbed radiations and subsequently in reduced photosynthetic rate [75]. The interactions between plants and different types of pollutants were investigated by many authors: most of the studies on the influence of environmental pollution focusing on physiological and ultrastructural aspects [76], [77]. Studies concerning the anatomy of the vegetative organs under conditions of pollution have been also carried out [78], [79], [80], [81], [82]. Dineva [83] and Tiwari et al. [75] recorded a reduction of leaf area and petiole length under pollution

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications stress conditions. Previous researches reported significant reduction in different leaf variables in the polluted environment in comparison with clean atmosphere [84]. In their study on Platanus acerifolia showed changes in leaf blade and petiole size in the polluted air. Significant reduction in length and area of leaflets and length of petiole of *Guaiacum officinale* of polluted plants was recorded. Reduction in the dimension of a leaf blade of five tree species in the vicinity of heavy dust and SO₂ pollution was also observed [84]. Significant effects of automobile exhaust on the phenology, periodicity and productivity of roadside tree species was also reported [85]. The decrease in leaf area in drought stress had been observed because of tolerance of water content of tissue possible by the decrease in leaf area [86]. Increase in length, breadth of leaflets and decrease in the area of the leaf had demonstrated in leaves of Albizia lebbeck under the stress of air pollution [87], [88]. Moreover, a study on leaves of Callistemon citrinus planted in industrial area clears that length, width of leaf and also leaf area decreased [88]. Cassia siamea plants growing at two different sites (polluted and non-polluted) on two important roads of Agra city exhibited significant differences in their flowering phonology and floral morphology [89]. Researchers showed that stimulation of photosynthetic rates in elevated CO₂ was nullified by decreased total leaf area [90]. Totally describe of air pollution is related to the morphology of area of leaf visible damage including a reduction of leaf area, changes in morphology as compare to unpolluted condition, necrosis and chlorosis. Naido and Chricot [91] showed that by the effect of air pollutant exchange of gases on the area of a leaf of Avicenia marina decreased. One way to increase tolerance in contrast with stress is to balance the water content of tissue by decrease the leaf area [86]. It seems that these species use this way as a defense mechanism.

Dust Effect on photosynthesis through deposition

The substantial effect of coal dust deposition on plant leaf surfaces probably lies thereby in a reduction of the photosynthetic product [91], [92]. Krajickova and Mejstrik [93] found that particulate matters from a coal-fired power plant affected photosynthesis of Calamagrostis epigejos and Hypericum perforatum but the stomata were rarely blocked. They suggested that the dust might act directly on the guard cells, though the mechanisms for this effect remain uncertain until now. After dust deposition on the leaves, Rhododendron catawbiense exhibited an increased absorption in the infrared spectrum and a reduced reflection and transmission of radiation [94]. Deposition of smaller particle sizes leads to stranger reduction of photosynthesis than with coarse particles [95]. This effect is presumably due to the closer lining of dust particles on leaf surface resulting in a greater shading effect of photosynthetically active radiation. In experiments, where the dust of different particle sizes were applied electrostatically to Brassica plant leaves, no difference in the photosynthetic efficiency of the plant was found. This may be due to uniform particle distribution and a very thin layer of dust deposition. Plant reactions due to dust deposition species dependent. fluorescence of *Ilex* are For example, the chlorophyll

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications *rotunda* and *Ficus microcarpa* has grown close by a ceramic plant are less affected than for *Machilus chinensis*. Least changes of the photosystems II have been observed with *Ilex rotunda* [96]. The chlorophyll fluorescence data of the mangrove, *Avicennia marina*, indicated that leaves coated with dust exhibited significantly lower photosystem II (PS II) quantum yield, lower electron transport rate through PSII, and reduced quantum efficiency of PSII (F-v/F-m) [91]. Nanos and Ilias [97] are reporting similar results for olive trees exposed to cement dust. In their study, cement dust decreased leaf total chlorophyll content and chlorophyll *a*/chlorophyll *b* ratio. As a result, photosynthetic rate and quantum yield decreased. Long-term effects include a change in the leaf biochemistry and a possible increase in plant pathogen and *Phytophagous arthropods* incidence [98]. Taylor *et al.*, [98] reported leaf rolling and interveinal necrosis in *Phaseolus vulgaris* after deposition of cement. Sperber [99] already assumed that plants adapted to low light conditions are less affected by dust depositions than plants adapted to grow directly in the sun.

Dust may Effect on the Pigments Content

Air pollution stress leads to stomatal closure, which reduces CO₂ availability in leaves and inhibits carbon fixation. The net photosynthetic rate is a commonly used indicator of the impact of increased air pollutants on tree growth [100]. Plants that are constantly exposed to environmental pollutants absorb, accumulate and integrate these pollutants into their systems. It has reported that depending on their sensitivity level, plants show visible changes which would include alteration in the biochemical processes or accumulation of certain metabolites [101]. Sulphur dioxide (SO₂⁻), nitrogen oxides (NO_x) and CO₂ as well as suspended particulate matter. These pollutants, when absorbed by the leaves, may cause a reduction in the concentration of photosynthetic pigments viz., chlorophyll and carotenoids, which directly affected to the plant productivity [102]. A relationship between traffic density and photosynthetic activity, stomatal conductance, total chlorophyll content, and leaf senescence has been reported [103]. One of the most common impacts of air pollution is the gradual disappearance of chlorophyll and concomitant yellowing of leaves, which may be associated with a consequent decrease in the capacity for photosynthesis [104]. Chlorophyll is found in the chloroplasts of green plants and is called a photoreceptor. Chlorophyll itself is actually not a single molecule but a family of related molecules, designated as chlorophyll "a", "b", "c" and "d". Chlorophyll "a" is the molecule found in all plant cells and therefore its concentration is what is reported during chlorophyll analysis [105]. Chlorophyll is the principal photoreceptor in photosynthesis, the light-driven process in which carbon dioxide is "fixed" to yield carbohydrates and oxygen. When plants are exposed to the environmental pollution above the normal physiologically acceptable range, photosynthesis gets inactivated. The distribution of plant diversity is highly dependent on the presence of air pollutants in the ambient air and sensitivity of the plants. Chlorophyll measurement is an important tool to evaluate the effects of air pollutants on plants as it plays an important role in plant metabolism and any

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications reduction in chlorophyll content corresponds directly to plant growth [102]. Chlorophyll is an index of productivity of the plant. Whereas certain pollutants increase the total chlorophyll content, others decrease it [101]. Changes in concentration of pigments were also determined in leaves of six tree species exposed to air pollution due to vehicle emissions [102]. The shading effects due to deposition of suspended particulate matter on the leaf surface might be responsible for this decrease in the concentration of chlorophyll in a polluted area. It might clog the stomata thus interfering with the gaseous exchange, which leads to an increase in leaf temperature which may consequently retard chlorophyll synthesis. Dusted or encrusted leaf surface is responsible for reduced photosynthesis and thereby causing a reduction in chlorophyll content [102]. A considerable loss in total chlorophyll, in the leaves of plants exposed to pollution supports the argument that the chloroplast is the primary site of an attack by air pollutants such as SO₂ and NO_x. Air pollutants make their entrance into the tissues through the stomata and cause partial denaturation of the chloroplast and decrease pigment contents in the cells of polluted leaves. A high amount of gaseous SO₂ causes destruction of chlorophyll [106]. Several pieces of research have recorded reduction in chlorophyll content under air pollution [75], [102], [104], [105], [106]. On the contrary, several pieces of research have exhibited an increase in chlorophyll content under air pollution, such as Tripathi and Gautam [106] reported that Mangifera indica leaves subjected to air pollution showed an increase (12.8%) in chlorophyll content [106]. Agbaire and Esiefarienrhe [101] in a study have demonstrated that plants from experimental site contain more chlorophyll compared with those from the control. Increase in the content of chlorophyll a, chlorophyll b, total chlorophyll, and carotenoid in Albizia lebbeck and Callistemon citrinus, has been reported by Seyyednejad et al., [87]. Investigation proved that chlorosis, is the first indicator of Flour effect on the plant [107]. Yun [108] showed the reduction in photosynthesis because of the PSII function damage, in sensitive species of tobacco. Carotenoids exist in plasma of plant tissues, photosynthetic or non-photosynthetic; the function of carotenoids in chloroplasts is as pigments to capture the light. But probably, the more important role is in protecting the cells and live organisms encounter with damage of free radical oxidative [109]. Plants fumigated with 40, 80 and 120 ppbv concentrations of O₃ exhibited a significant reduction in total chlorophyll content, RuBP carboxylase activity and net photosynthesis [110]. Carotenoids protect photosynthetic organisms against potentially harmful photoxidative processes and are essential structural components of the photosynthetic antenna and reaction center [102]. Carotenoids are a class of natural fat-soluble pigments found principally in plants, algae, and photosynthetic bacteria, where play a critical role in the photosynthetic process. They act as accessory pigments in higher plants. They are tougher than chlorophyll but much less efficient in light gathering, help the valuable but much fragile chlorophyll and protect chlorophyll from photoxidative destruction [105]. In a study conducted by Joshi and Swami [104] on Eucalyptus cirtiodora subjected to air

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications pollution, highly reduced carotenoid contents were observed and in another study by the same group reported that the carotenoids concentration was reduced due to vehicular emission [102]. Several researchers have reported reduced carotenoid content under air pollution [75], [105], [106].

Dust Interference with Stomata functions

The effects are devastating in an arid environment, because even drought-tolerant crops, if wind stressed, show an increased rate of water loss due to increased stomatal conductance, abrasion of the vapor barrier and an increase of transpiration [110], [111]. A positive effect of the vegetation to the environment is its air filtering ability: dust particles in the atmosphere get deposited on the leaves, which improves the air quality. The leaves' ability to act as dust receptors depends upon their surface geometry, orientation, epidermal and cuticle features, leaf pubescence and plant height [112]. However, at the same time, dust on the leaf surface affects plant growth and development. For example, dust deposits on the leaves can alter their optical properties, especially the surface reflectance in the visible and near-infrared range of the wavelength [113]. Dust particulate matters deposition can cause blockage of the stomata on the upper surfaces of the vegetation under natural condition and to a smaller extent at the lower surfaces. According to Krajickova and Mejstrik [93], the stomatal diameters of most plants usually range from 8-12 µm. Therefore, dust particle size is an important criterion for a possible leave penetration. Particles with PM₁₀ and smaller sizes can theoretically interfere with stomatal functions. Clogging the leaf stomata lowers the rate of transpiration and carbon assimilation, which finally causes a significant reduction in the photosynthesis rate. Dust affect less plant, which exhibits physical protection structures such as trichomes compared to plants without such physical barriers. Cornisch et al., [114] concluded that an increase in yield is associated with the stomatal conductance in Pima cotton (Gossypium barbadense L.). Stomatal conductance depends on environmental factors, the position at the canopy and age of the leaves [115]. Coal dust significantly reduced carbon dioxide exchange of upper and lower leaf surfaces of the mangrove, Avicennia marina by 17-39%, whereby the reduction was generally greater on the lower leaf surface, which has the dense mat of trichomes and salt glands [91]. Nano and Ilias [97] describe a decrease of the stomatal conductance to H₂O and CO₂ in Olea europaea exposed to cement dust, resulting in a reduced productivity of olive trees. Hirano et al., [64] found that dust decreased stomatal conductance of cucumber and kidney beans in the light, but increased it in the dark by plugging the stomata, when the stomata were open during dusting. When the dust of smaller size particles was applied to the plants, the effect was greater. However, the effect was negligible by closed stomata during dusting. Fluckiger et al., [116] found, that 1 mg cm⁻¹ of silica dust was necessary to cause a decrease in stomatal diffusive resistance in *Populus tremula*, but only 0.5 mg cm⁻² was necessary to cause an increase in leaf temperature. Metabolic functions in plants operate only in a certain optimal

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications temperature range. If a leaf heats up above 34°C, photosynthetic enzymes begin to denature and the leaf cannot perform its normal function. For example, Jiao and Grodzinski [117] describe an inhibition of photosynthetic export in *Salvia splendens* above 35°C in both photorespiratory conditions, whereby photosynthesis only under photorespiratory conditions was inhibited. Sucrose and raffinose but not stachyose accumulated in the leaf at 40°C. Plants react in this situation with an increase in transpiration to lower leaf temperatures.

Dust interaction with the Cuticle

Ulrichs *et al.*, [38] and Majumder *et al.*, [118] used silica dust applied to *Brassica* leaf surfaces as insecticides. The dust had strong lipophilicity and weak hydrophobic characteristics. Thereby, the dust got physically absorbed on the surface waxes of the leaves causing an irreversible damage resulting in a reduced photosynthetic rate. Bacic *et al.*, [119] made comparisons between the surfaces of *Pinus halepensis* needles from a site with relatively clean air and one near to a cement factory in Croatia. Induced changes in the appearance and quantity of surface wax were recorded only for the samples collected near to the cement factory. In particular, crystalline wax in suprastomatal cavities appeared to coalesce and subsequently additional amorphous wax formed around the rim of the stoma. However, this effect depends on physico-chemical properties of the dust particulates and environmental conditions. Ulrichs *et al.*, [120] showed that application of fly ash on leaves did not interact with the leaves in open fields. In open field conditions, small size particles drift easily from the leaf surface via air movement and precipitation. Therefore, dusts impair directly neither leaf surfaces nor photosynthesis significantly.

Dust effect on Sugar

Soluble sugar is an important constituent and source of energy for all living organisms. Plants manufacture this organic substance during photosynthesis and breakdown during respiration [106]. Tripathi and Gautam [106], in their study, revealed a significant loss of soluble sugar in all tested species at all polluted sites. The concentration of soluble sugars is indicative of the physiological activity of a plant and it determines the sensitivity of plants to air pollution. Reduction in soluble sugar content in polluted stations can be attributed to increased respiration and decreased CO₂ fixation because of chlorophyll deterioration. It has been mentioned that pollutants like SO₂, NO₂, and H₂S under hardening conditions can cause more depletion of soluble sugars in the leaves of plants grown in the polluted area. The reaction of sulfite with aldehydes and ketones of carbohydrates can also cause a reduction in carbohydrate content [106]. Some researchers showed that Concentrations of total and soluble sugars decreased significantly in the sensitive trees to the air pollution. In damaged *Quercus cerris* leaves the decrease in concentrations of sugars was higher in September. The decrease in total sugar content of damaged leaves probably corresponded with the photosynthetic inhibition or stimulation of respiration rate [121]. Furthermore, an increase in the amount of soluble sugar is a protection mechanism of leaves

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications it has been shown in Pinto bean in exposure with different concentration of ozone [122]. Following ozone exposure, soluble sugars in pine needle decreased [123]. Subsequently, they increased, frequently in association with foliar injury [122], [124]. The increase of soluble sugars was also observed following chronic exposure [124]. The increase in soluble sugar was reported in *Albizia lebbeck* and *Callistemon citrinus* grown in the industrial land [88]. Investigations revealed that the more resistant species plants to the air pollution as compared to sensitive species showed more concentration of soluble sugar [125], [126].

Dust effect on Proline

Some workers have been published the increase in free proline content in response to various environmental stresses in plants [127]. Typical environmental stress (high and low temperature, drought, air, and soil pollution) can cause excess Reactive Oxygen Species (ROS) in plant cells, which are extremely reactive and cytotoxic to all organisms [127]. High exposure to air pollutants forces chloroplasts into an excessive excitation energy level, which in turn increases the generation of ROS and induces oxidative stress [100]. The deleterious effects of the pollutants are caused by the production of Reactive Oxygen Species (ROS) in plants, which cause the peroxidative destruction of cellular constituents [75]. It has been reported that proline act as a free radical scavenger to protect plants away from damage by oxidative stress. Although the scavenging reaction of ROS with other amino acids, such as tryptophan, tyrosine, histidine, etc. are more effective compared with proline, proline is of special interest because of its extensive accumulation in plants during environmental stress [128]. Tankha and Gupta [129] showed the increase in the content of proline with increasing SO₂ concentration. According to existence of SO₂ and CO in the industrial area as the result of chemical activities, these results probably indicate that it has been clearly inconceivable to designate a harmless threshold toxic SO₂ concentration for level of particular species since other environmental factors during pollution profoundly affect the degree of damage [87]. A significant increase in the content of proline in Albizia lebbeck grown in the polluted area has been reported the concentration of proline increased in leaves of Callistemon citrinus planted round petrochemical site in comparison with control site [88]. Proline is a universal osmolyte accumulated in response to stress and may have a role in plant defense reactions [130]. Obviously proline has main role in protection in different kinds of stress. Accumulation of proline in plants is a physiological response to osmotic stress [131]. The effects of pollutants on plants include pigment destruction, depletion of cellular lipids and peroxidation of polyunsaturated fatty acid [75]. There appears to be a relationship between lipid peroxidation and proline accumulation in plants subjected to diverse kinds of stress, for example, proline accumulation in leaves of plants exposed to SO₂ fumigation [129], heavy metals [128] and salt [132] stress has been reported [128], [129], [132].

www.rjlbpcs.com

Dust effect over the soil

Dust particulates drift resulting from agricultural liming and fertilization can have an eutrophication effect on nearby soils. The best-studied dust particulates depositions are for coal fly ash from power plants. For decades, numerous researchers have looked into the possible use of coal fly ash in agriculture [133], [134], [135]. Hard coal fly ash is a small less, grey, fine-grained and powdery substance, which consists mainly of spherical, glassy particles. Main components of coal fly ash are SiO₂, Al₂O₃, and Fe₂O₃. Both Logan and Harrison [136] and Wong [137] described coal fly ash as rich in calcium and magnesium oxide and thus explaining the high pH value observed by others. Coal fly ash contains polychlorinate biphenyls, polycyclic aromatic hydrocarbons, and various metals in the mg per kg range. In various investigations, fly ash as substrate was used and data were interpreted from the viewpoint of plant nutrition [138], [139], [140]. Generally, changes in soil chemistry after dust particulates depositions may be most important for long-term effects on plants [141].

Soil nutritional value

Plants use inorganic minerals for nutrition. Many factors influence nutrient uptake for plants. Ions can be readily available to roots or could be "tied up" by other elements or the soil itself. Soil alkaline or acidic in pH makes minerals unavailable to plants. The optimal soil pH ranges for most crop plants between 5.5 to 6.2 or slightly acidic. This creates the greatest average level for availability for all essential plant nutrients. Therefore, extreme fluctuations in pH can cause deficiency or toxicity of nutrients. Cawse et al., [142] found that rainfall around a cement plant in South Wales was high in phosphorus and vanadium and had a pH in the alkaline range. Garden cress was relatively insensitive to pH changes [143], [120]. Theis and Wirth [144] found that major components of coal fly ash particulates were Al. Fe and Si with smaller concentrations of Ca, K, Na, Ti, S and numerous trace elements. Some of those elements like Ca, Fe, Mg and K are required for plant growth [145], [146]. Some others like Be, Se and Mo can be toxic. Generally, coal fly ash is not an optimal source of phosphorus since it was found to be inferior to monocalcium phosphate [147]. However, Ca²⁺ and Mg²⁺ can increase plant growth, as shown for legumes [133], [134]. Next, to the nutritional value, dust particulates can have negative effects on the soil nutritional value. As for example, alarming concentrations of lead were found in the dust of densely populated urban areas and in water and land of various areas near the industrial waste disposals [148]. Plants absorb lead and accumulation of this metal is reported for roots, stems, leaves, root nodules, seeds etc. [149]. Furthermore, the lead content of plant tissues increases with the increase of exogenous lead level. Lead affects plant growth and productivity, whereby the magnitude of the effect depends on the plant species. Photosynthesis has been found to be one of the most sensitive plant processes and the effect of the metal is multifacial. Lead also inhibits Nitrogen fixation and NH₄ assimilation in the root nodules. It appears that toxic effect of this

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications metal is primarily at the physiological level [148]. Warambhe et al., [150], had evidence that high coal fly ash depositions result in high soil pH values and phytotoxic boron contents in these areas that disturb plant growth. Only after sufficient precipitation, the phytotoxic characteristics of the substrates with very high coal fly ash content decrease. Other researchers claimed that higher boron contents in coal fly ash have soil-ameliorating characteristics [137]. Cline *et al.*, [151] showed that yields of soybeans increased up to 35% by coal fly ash applications on sandy and clay soils in the south of Ontario. On the yield of corn, coal fly ash had, however, no effects.

Soil texture and density

Normally, dust particulates deposition makes only a fraction of the topsoil volume. Therefore, a change of the physical structure of the soil is very unlikely. This is, of course, different dust particulates are collected and artificially deposited or mixed in the soil. For example, soil properties are influenced by coal fly ash application [152]. Here the physical and chemical properties of soil vary according to the original properties. Several researchers described the role of increasing coal fly ash contents in the soil with positive effects on the water holding capacity in sandy [153], [154], and coarse-grained [155] soils.

Impact of Dust on Plant Communities

Some of the earliest references regarding to dust influences on plant community structures date back to 1910. Parish [156] was interested in the shrub and grassland vegetation in California near cement factories. He found a shift in the vegetation community close to some cement factories [157]. Lotschert and Kohm [158] reported bark pH and Ca² content changes in the bark of trees after long-term dust exposure. Since water stress is one of the major urban stressors for trees such findings can help to select plant species and varieties in anthropogenically determined systems.

2. CONCLUSION

Present work provides basic information about the variation in dust particles accumulation of plant species. Variation of dust pollutions positively shows the impacts on plants species with the alteration of morphological, biochemical, epidermal, stomata functions, soil conditions, and soil nutritions and also species communities etc. Deposition of quarry dust has resulted in the reduction in the leaf area and chlorophyll content and also in the stunted growth. In this article, we summarized the influences of dust pollution on various parameters of vegetation.

ACKNOWLEDGEMENT

One of the authors YG thankful to the DST (Inspire Program), New Delhi for providing financial support.

CONFLICT OF INTEREST

The authors have no conflict of interest.

- 1. Anderson JO, Thundiyil JG and Stolbach A. Clearing the air: a review of the effects of particulate matter air pollution on human health. J. Med. Toxicol. 2012; 8: 166-175.
- 2. Kan H, Chen R and Tong S. Ambient air pollution, climate change, and population health in China. Environ. Int. 2012; 42: 10-19.
- 3. Lovett GM, Tear TH, Evers DC, Findlay SEG, Cosby BJ, Dunscomb JK, Driscoll CT and Weathers KC. Effects of air pollution on ecosystems and biological diversity in the eastern United States. Ann. N.Y. Acad. Sci. 2009; 1162: 99-135.
- 4. Walsh M and Shah JJ. Clean Fuels for Asia: Technical Options for Moving Toward Unleaded Gasoline and Low-Sulfur Diesel. World Bank Technical Paper No. 377. The World Bank, Washington D.C., 1997.
- 5. Ning Z and Sioutas C. Atmospheric processes influencing aerosols generated by combustion and the inference of their impact on public exposure: A review. Aerosol Air Qual. Res. 2010; 10: 43-58.
- 6. Cao L, Appel E, Hu S, Yin G, Lin H and Rosler W. Magnetic response to air pollution recorded by soil and dust-loaded leaves in a changing industrial environment. Atmos. Environ. 2015; 119: 304-313.
- 7. World Health Organization (WHO), Health aspects of air pollution with particulate matter, ozone and nitrogen dioxide. Report 2003 on a WHO working group. Bonn; 2003, http://www.euro.who.int/_data/asset s/pdf_file/0005/112199/E79097.pdf.
- 8. World Health Organization (WHO), the World Health Report 2002: Reducing Risks, Promoting Healthy Life. WHO, Geneva, 2002.
- 9. Karanasiou A, Amato F, Moreno T, Lumbreras J, Borge R, Linares C, Boldo E, Alastuey A and Querol X. Road dust emission sources and assessment of street washing effect. Aerosol Air Qual. Res. 2014; 14: 734-743.
- 10. Upadhyay N, Clements AL, Fraser MP, Sundblom M, Solomon P and Herckes P. Sizedifferentiated chemical composition of re-suspended soil dust from the Desert Southwest United States. Aerosol Air Qual. Res. 2015; 15: 387-398.
- 11. Begum BA, Biswas SK, Pandit GG, Saradhi IV, Waheed S, Siddique N, Seneviratne MCS, Cohen DD, Markwitz A and Hopke PK. Long-range transport of soil dust and smoke pollution in the South Asian region. Atmos. Pollut. Res. 2011; 2: 151-157.
- 12. Schelle-Kreis J, Sklorz M, Herrmann H and Zimmermann R. Sources, occurrences, compositions – Atmospheric aerosols. Chemie in Unserer Zeit. 2007; 41: 220-230.
- 13. Tong CH, Blanco M, Goddard WA and Seinfeld JH. Secondary organic aerosol formation by heterogeneous reactions of aldehydes and ketones: A quantum mechanical study. Environmental Science and Technology. 2006; 40: 2333-2338.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications
14. Gibson ER, Gierlus KM, Hudson PK and Grassian VH. Generation of internally mixed insoluble and soluble aerosol particles to investigate the impact of atmospheric aging and heterogeneous processing on the CCN activity of mineral dust aerosol. Aerosol Science and Technology. 2007; 41: 914-924.

- Kulshrestha MJ, Kulshrestha UC, Parashar DC and Vairamani M. Estimation of SO₄ contribution by dry deposition of SO₂ onto the dust particles in India. Atmos. Environ. 2003; 37: 3057- 3063.
- 16. Gupta GP, Kumar B, Singh S. and Kulshrestha UC. Urban climate and its effect on biochemical and morphological characteristics of Arjun (*Terminalia arjuna*) plant in National Capital Region Delhi. Chem. Ecol. 2015; 31: 524-538.
- Gupta GP, Singh S, Kumar B and Kulshrestha UC. Industrial dust sulphate and its effects on biochemical and morphological characteristics of Morus (*Morus alba*) plant in NCR Delhi. Environ. Monit. Assess. 2015; 187: 67.
- 18. Sheppard LJ. Causal mechanisms by which sulphate, nitrate and acidity influence frost hardiness in red spruce: Review and hypothesis. New Phyto. 1994; 127: 69- 82.
- Grantz DA, Garner JHB and Johnson DW. "Ecological effects of particulate matter". Environ. Int. 2003; 29: 213-239.
- 20. Buchner P, Takahashi HM and Hawkesford J. Plant sulphate transporters: Co-ordination of uptake, intracellular and long-distance transport. J. Exp. Bot. 2004; 55: 1765-1773.
- 21. Vardak E, Cook CM and Lanaras T. Effect of dust from a limestone quarry on the photosynthesis of *Quercus coccifera*, and evergreen Sclerophyllous shrub. Bull. Environ. Contam.Toxicol. 1995; 54: 414-419.
- 22. Auerbach NA, Walker MD and Walker DA. Effects of roadside disturbance on substrate and vegetation properties in arctic tundra. Ecol. Appl. 1997; 7: 218-235.
- Hope AS, Fleming JB and Stow DA. Tussock tundra albedos on the north slope of Alaska: Effects of illumination, vegetation composition, and dust deposition. J. Appl. Meteorol. 1991; 30: 1200-1206.
- 24. Gupta GP, Kumar B, Singh S and Kulshrestha UC. Deposition and Impact of Urban Atmospheric Dust on Two Medicinal Plants. Aerosol and Air Quality Research. 2016; 16: 2920-2932.
- 25. Spatt PD and Miller MC. Growth conditions and vitality of Sphagnum tundra community along the Alaska Pipeline Haul Road. ARCTIC. 1981; 34: 48-54.
- 26. Spencer S and Tinnin R. Effects of coal dust on plant growth and species composition in an arid environment. J. Arid Environ. 1997; 37: 475-485.
- 27. Walker DA and Everett KR. Road dust and its environmental impact on Alaskan taiga and tundra. Arct. Alp. Res. 1987; 19: 479-489.

Kameswaran et al RJLBPCS 2019www.rjlbpcs.comLife Science Informatics Publications28. Sharifi MR, Gibson AC and Rundel PW. Surface dust impacts on gas exchange in Mojave

- Desert shrubs. J. Appl. Ecol. 1997; 34: 837-846.
- 29. Wijayratne UC, Scoles-Scilla SJ and Defalco LA. Dust deposition effects on growth and physiology of the endangered *Astragalus Jaegerianus* (Fabaceae)". Madroño. 2009; 56: 81-88.
- 30. Farmer AM. The effects of dusts on vegetation A review. Environ. Pollut. 1993; 79: 63-75.
- 31. Beckett KP, Freer-Smith PH and Taylor G. Particulate pollution capture by urban trees: Effect of species and wind speed. Global Change Biol. 2000; 6: 995-1003.
- 32. Fennelly PF. Primary and secondary particulates as pollutants. Journal of the Air Pollution Control Association. 1975; 25: 397-704.
- Brook JR, Poirot RL, Dann TF, Lee PKH, Lillyman CD and Ip T. Assessing sources of PM2.5 in cities influenced by regional transport. Journal of Toxicol. Environ. Health. 2007; 70: 191-199.
- 34. Gilmour MI, McGee J, Duvall RM, Dailey L, Daniels M, Boykin E, Cho SH, Doerfler D, Gordon T and Devlin RB. Comparative toxicity of size-fractionated airborne particulate matter obtained from different cities in the United States. Inhalation Toxicology. 2007; 19: 7-16.
- 35. Langner M. Staubumsatz in verkehrsexponierten Baumkronen und Partikelverteilung in stadtischen Grunflachen. Berliner Geographische Arbeiten. 2007; 109, 1-12.
- Mewis and Ulrichs Ch. Effects of diatomaceous earth on water content of *Sitophilus granaries* (L.) (Col.: Curculionidae) and its possible use in stored product protection. Journal of Applied Entomology. 2001; 125: 351-360.
- 37. Mewis and Ulrichs Ch. "Action of amorphous diatomaceous earth against different stages of the stored product pests *Tribolium confusum*, *Tenebrio molitor*, *Sitophilus granaries* and *Plodia interpunctella*". Journal of Stored Products Research. 2001; 37: 153-164.
- 38. Ulrichs Ch, Krause F, Rocksch T, Goswami A and Mewis I. "Electrostatic application of inert silica dust based insecticides to plant surfaces". Communications in Agricultural and Applied Biological Sciences, Ghent University. 2006; 71: 171-178.
- 39. Darley EF. Studies on the effect of cement-kiln dust. Journal of the Air Pollution Control Association. 1966; 16: 145-150.
- 40. Brown J, Berg R (Eds) Environmental Engineering and Ecological Baseline Investigations along the Yukon River-Purdhoe Bay Haul Road, US Army Cold Regions Research and Engineering Laboratory", CRREL Report 80-19. 1980; Pp: 101-128.
- 41. Sabin LD, Lim JH, Venezia MT, Winer AM, Schiff KC, and Keith DS. "Dry deposition and resuspension of particles-associated metals near a freeway in Los Angeles". Atmos. Environ. 2006; 40: 7528-7538.
- 42. Biasioli M, Barberis R and Ajmone-Marsan F. "The influence of a large city on some soil properties and metals content". Sci. Total Environ. 2006; 356: 154-164.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications
43. Mielke HW, Laidlaw MA and Gonzales C. "Lead (Pb) legacy from vehicle traffic in eight California urbanized areas: Continuing influence of lead dust on children's health". Sci. Total Environ. 2010; 408: 3965-3975.

- 44. Kong SF, Lu B, Ji YQ, Zhao XY, Chen L, Li ZY, Han B and Bai ZP. "Levels, risk assessment and sources of PM10 fraction heavy metals in four types dust from a coal-based city". Microchem. J. 2011; 98: 280-290.
- 45. Bell ML, Morgenstern RD and Harrington W. Quantifying the human health benefits of air pollution policies: review of recent studies and new directions in accountability research. Environ. Sci. Policy. 2011; 14: 357-368.
- 46. Chen X, Xia X, Zhao Y and Zhang P. Heavy metal concentrations in roadside soils and correlation with urban traffic in Beijing, China. J. Hazard. Mater. 2010; 181: 640-646.
- 47. Ali H., Khan, E. and Sajad, M.A. Phytoremediation of heavy metals Concepts and applications. Chemosphere. 91: 869-881, 2013.
- 48. Ram SS, Kumar RV, Chaudhuri P, Chanda S, Santra SC, Sudarshan M and Chakraborty A. Physicochemical characterization of street dust and re-suspended dust on plant canopies: An approach for finger printing the urban environment. Ecol. Indic. 2014; 36: 334-338.
- 49. Norouzi S, Khademi H, Cano AF and Acosta JA. Using plane tree leaves for biomonitoring of dust borne heavy metals: A case study from Isfahan, Central Iran. Ecol. Indic. 2015; 57: 64-73.
- 50. Freer-Smith PH, Sophy H and Goodman A. The uptake of particulates by urban woodland: Site description and particulate composition. Environ. Pollut. 1997; 95: 27-35.
- Prusty BAK, Mishra PC and Azeez PA. Dust accumulation and leaf pigment concentration in vegetation near the national highway at Sambalpur, Orissa, India. Ecotoxicol. Environ. Saf. 2005; 60: 228-235.
- 52. Nowak DJ, Crane DE and Stevens JC. Air pollution removal by urban trees and shrubs in the United States. Urban Forestry and Urban Greening. 2006; 4: 115-123.
- 53. Jamil S, Abhilash PC, Singh A, Singh N and Behl HM. Fly ash trapping and metal accumulating capacity of plants: Implication for green belt around thermal power plants. Landscape Urban Plann. 2009; 92: 136-147.
- 54. Qiu Y, Guan D, Song, W and Huang K, Capture of heavy metals and sulfur by foliar dust in urban Huizhou, Guangdong Province, China. Chemosphere. 2009; 75: 447-452.
- 55. Hofmann H, Bartholomeus K, Calders SV, Wittenberghe K, Wuyts and Samson R. On the relation between tree crown morphology and particulate matter deposition on urban tree leaves: A ground-based LiDAR approach. Atmos. Environ. 2014; 99: 130-139.
- MOELLER D, Luft. Chemie, Physik, Biologie, Reinhaltung, Recht. Berlin, de Gruyter, 2003. Mit 168 Abb. u. 178 Tab. 23, 750 S. Farb. ill. OPbd. - Sehr gutes Ex.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications
57. Wolf-Benning U. Kleinraumige und zeitliche Variabilitat von Feinstaub und Grobstaub sowie Stickstoffdioxid in Berlin. Berliner Geographische Arbeiten. 2006; 105: 130.

- 58. Belot Y, Baille A and Delmas JL. Modele numerique de dispersion des pollutants atmospheriques en presence de couverts vegetaux. Atmospheric Environnment. 1976; 10: 89-98.
- 59. Chamberlain AC, Transport of Lycopodium spores and other small particles to rough surfaces. Proceedings of the Royal Society of London Ser. A. 1967; 296: 45-70.
- 60. Duggar BM and Cooley JS. The effect of surface films and dusts on the rate of transpiration. Annals of the Missouri Botanical Garden. 1914; 1: 1-22.
- 61. Sree Rangasami SR and Jambulingam R. Cement dust pollution on maize crop. Madras Agricultural Journal. 1973; 60: 1310-1313.
- Roberts JW, Watters HA, Mangold CA and Rossano AT. Cost and benefits of road dust control in Seattle's industrial valley. Journal of the Air Pollution Control Association. 1975; 25: 948-952.
- 63. Oblisami G, Pathmanabhan G and Padmanabhan C. Effect of particulate pollutants from cement-kilns on cotton plants. Indian Journal Air Pollution Control. 1978; 1: 91-94.
- 64. Hirano T, Kiyota M and Aiga I. Physical effects of dust on leaf physiology of cucumber and kidney bean plants. Environmental Pollution. 1995; 89: 255-261.
- 65. Borka G. The effect of cement dust pollution on growth and metabolism of *Helianthus annus*. Environmental Pollution (Ser.A). 1980; 22: 75-89.
- 66. Steinhubel G. and Halas L. Poruchy v tvorbe susiny pri zvysenych teplotach vyvolanych v listoch drevin prasnou imisiou. Lesnicky Casopis. 1967; 13: 365-383.
- 67. Guggenheim R, Fluckiger W, Fluckiger-Keller H and Oertli JJ. Pollution of leaf surfaces in the vicinity of a motorway. Berichte Umweltbundesamt. 1980; 79: 462-468.
- 68. Eveling DW. Effects of spraying plants with suspensions of inert dusts. Annals of Applied Biology. 1969; 64: 139-151.
- 69. Mage D, Ozolins G, Peterson P, Webster A, Orthofer R, Wandeweerd V and Gwynne M. Urban air pollution in megacities of the world. Atmospheric Environ. 1996; 30: 681-686.
- 70. Davison, A. W. & Blakemore, J. (1976). Factors determining fluoride accumulation in forage. Pp.17-30 in Effects of air pollutants on plants (Ed. T.A. Mansfied). Cambridge University Press, Cambridge, UK: Pp: 209.
- Chaphekar SB, Boralkar DB and Shetye RP. Plants for air monitoring in industrial area. In: Furtado JI (ed.) Tropical Ecology and Development. I.S.T.E. Kuala Lampur, 1980; pp. 669-675.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com

Life Science Informatics Publications

- 72. Chaturvedi RK, Prasad S, Rana S, Obaidullah SM, Pandey V and Singh H. Effect of dust load on the leaf attributes of the tree species growing along the roadside. Environ. Monit. Assess. 2013: 185: 383-391.
- 73. Ahmad MJ, Akhtar Z, Zahir A and Jamil A. Effect of cadmium on seed germination and seedling growth of four wheat (Triticum aestivum L.) cultivars. Pak. J. Bot. 2012; 44: 1569-1574.
- 74. Uaboi-Egbenni PO, Okolie PN, Adejuyitan OE, Sobande AO, and Akinyemi O. Effect of industrial effluents on the growth and anatomical structures of Abelmoschus esculentus (Okra). Afr. J. Biotechnol. 2009; 8: 3251-3260.
- 75. Tiwari S, Agrawal M and Marshall FM. Evaluation of ambient air pollution impact on carrot plants at a sub urban site using open top chambers. Environ. Monit. Assess. 2006; 119: 15-30.
- 76. Heumann HG. Ultrastructural localization of zinc in zinc-tolerant Armeria maritime sp. halleri by autometallography. Journal of Plant Physiology. 2002; 159:191-203.
- 77. Psaras GK and Christodoulakis NS. Air pollution affects on the ultrastructure of Phlomis fruticosa mesophyll cells. Bulletin of Environmental Contamination and Toxicology. 1987; 38: 610-617.
- 78. Alves ES, Baêsso Moura M and Domingos M. Structural analysis of Tillandsia usneoides L. exposed to air pollutants in São Paulo City-Brazil. Water Air and Soil Pollution. 2008; 189: 61-68.
- 79. Ahmad SH, Reshi Z, Ahmad J and Iqbal MZ. Morpho-anatomical responses of Trigonella foenum-graecum L. to induced cadmium and lead stress. Journal of Plant Biology. 2005; 48: 64-84.
- 80. Silva LC, Azevedo AA, Silva EAM and Oliva MA. Effects of simulated acid rain on the growth and anatomy of five Brazilian tree species. Australian Journal of Botany. 2005; 53:789-796.
- 81. Silva C, Oliva MA, Azevedo AA and Araújo JM. De Responses of Restinga plant species to pollution from an iron pelletization factory. Water Air and Soil Pollution. 2006; 175:241-256.
- 82. Verma RB, Mahmooduzzafar TO, Siddiqi and Iqbal M. Foliar Response of Ipomea pes-tigridis L. to Coal Smoke Pollution. Turkish Journal of Botany. 2006; 30:413-417.
- 83. Dineva SB. Comparative studies of the leaf morphology and structure of white ash Fraxinus americana L. and London plane tree Platanus acerifolia Wild growing in polluted area. Dendrobiology. 2004; 52: 3-8.
- 84. Jahan S and Iqbal MZ. Morphological and anatomical studies on leaves of different plants affected by motor vehicle exhausted. J. Islamic Acad. Sci. 1992; 5: 21-23.

Kameswaran et al RJLBPCS 2019

www.rjlbpcs.com

- Life Science Informatics Publications 85. Bhatti GH and Iqbal MZ. Investigations into the effect of automobile exhausts on the phenology, periodicity and productivity of some roadside trees. Acta Sociotatis Botanicorum Poloniae. 1988: 57: 395-399.
- 86. Hale MG, Orcutt DM and Thompson LK. The Physiology of Plants under Stress. 1st Edn., Wiley Interscience Publication, Wiley and Sons Inc., New York. 1987; 206.
- 87. Seyyednejad SM, Niknejad M and Yusefi M. Study of air pollution effects on some physiology and morphology factors of Albizia lebbeck in high temperature condition in Khuzestan. J. Plant Sci. 2009; 4: 122-126.
- 88. Seyyednejad SM, Niknejad M and Yusefi M. The effect of air pollution on some morphological and biochemical factors of Callistemon citrinus in petrochemical zone in South of Iran. Asian J. Plant Sci. 2009; 8: 562-565.
- 89. Chauhan SV, Chaurasia B and Rana A. Impact of air pollution on floral morphology of Cassia siamea Lamk. J. Environ. Biol. 2004; 25: 291-297.
- 90. Noormets A, McDonald EP, Dickson RE, Kruger EL, Sober A, Isebrands JG and Karnosky DF. The effect of elevated carbon dioxide and ozone on leaf- and branch-level photosynthesis and potential plant-level carbon gain in aspen. Trees-Structure Function. 2001; 15: 262-270.
- 91. Naido G and Chricot D. The effect of coal dusts on photosynthetic performance of mangrove, Avicenia marina in Richards bay, South Africa. Environ. Pollut. 2004; 127: 359-366.
- 92. Auclair D. Effects of dust on photosynthesis 2. Effects of particulate matter on photosynthesis of scots pine and poplar. Annales des sciences Forestieres. 1977; 34: 47-57.
- 93. Krajickova A and Mejstrik V. The effect of fly-ash particles on the plugging of stomata. Environmental Pollution. 1984. 36: 83-93,
- 94. Eller BM and Brunner U. Der Einfluss von Straßenstaub auf die Strahlungsabsorption durch Blatter. Archiv fur Meteorologie, Geophysik und Bioklimatologie, Serie B. 1975; 23: 137-146.
- 95. Hirano T, Kiyota M, Kitaya Y and Aiga I. The physical effects of dust on photosynthetic rate of plant leaves. Journal of Applied Meterology. 1990; 46: 1-7.
- 96. Wen E, Kuang Y and Zhou G. Sensitivity analyses of woody species exposed to air pollution based on eco-physiological measurements. Environmental Science and Pollution Research. 2004; 11: 165-170.
- 97. Nanos GD and Ilias IF. Effects of inert dust on olive (Olea europaea L.) leaf physiological parameters. Environmental Science and Pollution Research. 2007; 14: 212-214.
- 98. Taylor HJ, Ashmore MR and Bell JNB. Air Pollution Injury to Pollution, Imperial College for Environmental Technology, London, total 1986; Pp.33.
- 99. Sperber A. Auswirkungen von Staub auf Photosynthese und Stoffproduktion verschiedener Pflanzen. Inaugurat-Dissertation. University Bonn. 1975; Pp: 108.

Kameswaran et al RJLBPCS 2019 www

www.rjlbpcs.com

- Life Science Informatics Publications
- 100. Woo SY, Lee DK and Lee YK. Net photosynthetic rate, ascorbate peroxidase and glutathione reductase activities of *Erythrina orientalis* in polluted and non-polluted areas. Photosynthetica. 2007; 45: 293-295.
- 101. Agbaire PO and Esiefarienrhe E. Air Pollution Tolerance Indices (APTI) of some plants around Otorogun gas plant in Delta State, Nigeria. J. Applied Sci. Environ. Manage. 2009; 13: 11-14.
- 102. Joshi PC and Swami A. Air pollution induced changes in the photosynthetic pigments of selected plant species. J. Environ. Boil. 2009; 30: 295-298.
- 103. Honour SL, Bell JNB, Ashenden TW, Cape JN and Power SA. Responses of herbaceous plants to urban air pollution: Effects on growth, phenology and leaf surface characteristics. Environ. Pollut. 2009; 157: 1279-1286.
- 104. Joshi PC and Swami A. Physiological responses of some tree species under roadside automobile pollution stress around city of Haridwar, India. Environmentalist. 2007; 27: 365-374.
- 105. Joshi N., Chauhan A. and Joshi, P.C. Impact of industrial air pollutants on some biochemical parameters and yield in wheat and mustard plants. Environmentalist. 2009; 29: 398-404.
- 106. Tripathi A.K. and Gautam, M. Biochemical parameters of plants as indicators of air pollution. J. Environ. Biol. 2007; 28: 127-132.
- 107. Kendrick JB, Darly EF, Middieton JT and Paulus AO. Plant response to polluted air: Specific effects of air pollutants on plants vary according to plant species and modifying internal and external factors. California Agric. 1956; 10: 9-15.
- 108. Yun MH. Effect of ozone on CO₂ assimilation and PSII function in plants with contrasting pollutant sensitivities. Dissertation Abst. Int. 2007; Vol. 68, No. 10.
- 109. Fleschin S, Fleschin M, Nhta S, Pavel E and Mageara V. Free radicals mediate protein oxidation in biochemistry. Roum. Biotechnol. Lett. 2003; 5: 479-495.
- Chapla and Kamalakar JA. Metabolic responses of tropical trees to ozone pollution. J. Environ. Biol. 2004; 25: 287-290.
- Armbrust DV and Retta A. Wind and sandblast damage to growing vegetation. Ann. Arid Zone. 2000; 39: 273-284.
- Prajapati SK. Ecological effect of airborne particulate matter on plants. Environ. Skept. Crit. 2002; 1: 12-22.
- Prajapati SK and Tripathi BD. Seasonal variation of leaf dust accumulation and pigment content in plant species exposed to urban particulates pollution. J. Environ. Qual. 2008; 37: 865-870.

Kameswaran et al RJLBPCS 2019

www.rjlbpcs.com Life Science Informatics Publications 114. Cornish, Radin JW, Turcotte EL, Lu ZM and Zeiger E. Enhanced photosynthesis and stomatal conductance of Pima cotton (Gossypium harbadense L.) bred for increased yield. Plant Physiol. 1991; 97: 484-489.

- 115. Hitron O and Zur B. Differences in stomatal response within cotton canopy. Biotronic. 1990; 19:39-48.
- 116. Fluckinger W, Oertli JJ and Fluckiger H. Relationship between stomatal diffusive resistance and various applied particle sizes on leaf surface. Zeitschrift fur Pflanzenphysiologie. 1979; 91: 173-175.
- 117. Jiao J and Grodzinski B. The effect of leaf temperature and photorespiratory conditions on export of sugars during steady-state photosynthesis in Salvia splendens. Plant Physiology. 1996; 111: 169-178.
- 118. Majumder DD, Banerjee R, Ulrichs Ch, Mewis I and Goswami A. Nano-materials: Science of bottom-up and top-down. IETE Technical Review on "Nanotechnology education a paradigm shift". 2007; 24: 9-25.
- 119. Bacic T, Lynch AH and Cutler D. Reactions to cement factory dust contamination by Pinus halepensis needles. Environmental and Experimental Botany. 1999; 41: 155-166.
- 120. Ulrichs Ch, Dolgowski D, Mucha T, Reichmuth Ch and Mewis I. Insektizide und phytotoxische Wirkung von Steinkohlenglugasche. Gesunde Pflanzen. 2005; 57: 110-116.
- 121. Tzvetkova N and Kolarvo D. Effect of air pollution on carbohydrate and nutrients contents on some deciduous tree species. Bulg. J. Plant Physiol. 1996; 22: 53-56.
- 122. Dugger WM and Ting IP. Air pollution oxidant-their effects on metabolic processes in plants. Annu. Rev. Plant Physiol. 1970; 21: 215-234.
- 123. Wilkinson TG and Barnes RL. Effect of ozone on CO₂ fixation patterns in pine. Can. J. Bot. 1973; 9: 1573-1578.
- 124. Miller PR, Parmeter JR, Flick BH and Martinez CW. Ozone dosage response of Ponderosa pine seedlings. J. Air Pollut. Control Assoc. 1969; 19: 6-6.
- 125. Kameli A and Losel DM. Carbohydrate and water stress in wheat plants under water stress. New Phytologist. 1993; 125: 609-614.
- 126. Ludlow KF. Carbohydrates metabolism in drought stress leaves of pigeon pea (Cajanus cajan). J. Exp. Bot. 1993; 44: 1351-1359.
- 127. Levitt J. Respons of Plant to Environmental Stresses. 1st Edn., Academic Press, New York, 1972.
- 128. Wang F, Zeng B, Sun Z, and Zhu C. Relationship between proline and Hg²⁺-induced oxidative stress in a tolerant rice mutant. Arch. Environ. Contam. Toxicol. 2009; 56: 723-731.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com

- Life Science Informatics Publications 129. Tankha and Gupta RK. Effect of Water deficit and SO₂ on total soluble protein, nitrate reductase activity and free proline content in sun flower leaf. Biol. Planta. 1992; 34: 305-310.
- 130. Khattab H. The deffence mechanism of cabbage plant against phloem-sucking aphid (Brevicoryne brassicae L.). Aust. J. Basic Applied Sci. 2007. 1: 56-62.
- 131. Szekely G. The role of proline in Arabidopsis thaliana osmotic stress response. Acta Biologica Szegediensis. 2004. 48: 81-81.
- 132. Woodward AJ and Bennett IJ. The effect of salt stress and abscisic acid on proline production, chlorophyll content and growth of in vitro propagated shoots of Eucalyptus camaldulensis. Plant Cell Tissue Organ Cult. 2005; 82: 189-200.
- 133. Page AI, Elseewi AA and Straughan IR. Physical and Chemical properties from fly ash from coal-fired power-plants with special reference to environmental impacts. Residue Reviews. 1979; 71: 83-120.
- 134. Adriano DC, Page AL, Elseewi AA, Chang AC and Straughan I. Utilization and disposal of fly ash and other coal residues in terrestrial ecosystems: A review. Journal of Environmental Quality. 1980; 9: 333-344.
- 135. Maiti SS, Mukhopadhyay M, Gupta SK and Banerjee SK. Evaluation of fly ash as a useful material in agriculture. Journal of the Indian Society of Soil Science. 1990; 38: 342-344.
- 136. Logan TJ and Harrison BJ. Physical characteristics of alkaline stabilized sewage sludge (N-Viro soil) and their effects on soil physical properties. Journal of Environmental Quality. 1995; 4: 153-164.
- 137. Wong JWC and Su DC. Reutilization of coal fly ash and sewage sludge as an artificial soilmix: effects of pre-incubation on soil physic-chemical properties. Bioresource Technology. 1997; 59: 97-102.
- 138. Elseewi AA, Bingham FT and Page AL. Availability of sulphur in fly ash to plants. Journal of Environmental Quality. 1978; 7: 69-73.
- 139. Hill MF and Lamp CA. Use of pulverized fuel ash from Victorian brown coal as a source of nutrients for pasture species. Australian Journal of Experimental Agriculture. 1980; 20: 377-384.
- 140. Kalra MC, Jain, Joshi HC, Choudhary R, Harit RC, Vatsa BK, Sharma SK and Kumar SK. Fly ash as soil conditioner and fertilizer. Bioresource Technology. 1998; 64: 163-167.
- 141. Scheffer F, Prezmeck E and Wilms W. Untersuchungen uber den Einfluss von Zementofen-Flugstaub auf Boden und Pflanzen. Staub. 1961; 21: 251-254.
- 142. Cawse PA, Baker SJ and Page RA. The influence of particulate matter on rainwater acidity and chemical composition. U.K. Atomic Energy Authority Report R 13478, HMSO, London, 1989.

Kameswaran et al RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications
143. Grantzau E. Geeignete Keimpflanzen zur Prufung der Pflanzenvertrag lichkeit von Komposten, Substraten und Boden. Erich Schmidt, Berlin. 1997; Pp: 43.

- 144. Theis TI and Wirth JL. Sorptive behavior of trace metals on fly ash in aqueous systems. Environmental Science and Technology. 1977; 11: 1096-110.
- 145. Kachoo D, Dixit AK and Bali AS. Influence of crop residue, fly ash and varying starter dosages on growth, yield and soil characteristics in rice (*Oryza sativa*) - wheat (*Triticum aestivum*) cropping system under irrigated conditions of Jammu region. Indian Journal of Agricultural Science. 2006; 76: 3-6.
- 146. Inam A. Use of fly ash on turnip (*Brassica rapa* L.) cultivation. Pollution Research. 2007; 26: 39-42.
- 147. Martens DC. Availability of plant nutrients in fly ash. Compost Science. 1971; 12: 15-19.
- 148. Singh RP, Tripathi RD, Sinha SK, Maheshwari R and Srivastava HS. Response of higher plants to lead contaminated environment. Chemosphere. 1997; 34: 2467-2493.
- 149. Hevesy G. The absorption and translocation of lead by plants. Biochemical Journal. 1923;17: 439-445.
- 150. Warambhe PE, Kene DR, Thakre KK, Darange OG and Bhoyar VS. Evaluation of physicochemical properties of fly ash of thermal power station, Koradi (Nagpur) for its likely use in agriculture. Journal of Soils and Crops. 1993; 3: 75-77.
- 151. Cline JA, Bijl M and Torrenueva A. Coal fly ash as a soil conditioner for field crops in southern Ontario. Journal of Environmental Quality. 2000; 29: 1982-1989.
- 152. Grewal KS, Yadava PS, Mehta SC and Oswal MC. Direct and residual effect of fly ash application to soil on crop yield and soil properties. Crop.Res. 2001; 21: 60- 65.
- 153. Roberts TJ. The effect of land type and fine particle amendments on the emergence and growth of subterranean clover (*Trifolium subterraneum* L.) with particular reference to water relations. Australian Journal of Agricultural Research. 1966; 17: 657-672.
- 154. Campbell DJ, Fox WE, Aitken RL and Bell LC. Physical characteristics of sands amended with fly ash. Australian Journal of Soil Research. 1983; 21: 147-154.
- 155. Chang AC, Lund LJ, Page AL and Warnecke JE. Physical properties of fly ash amended soils. Journal of Environmental Quality. 1977; 6: 267-270.
- 156. Parish SB. The effect of cement dust on citrus trees. Plant World. 1910; 13: 288-291.
- 157. Sadhana Chaurasia, Ashwani Karwariya and Anand Dev Gupta, Impact of Cement Industry Pollution on Morphological Attributes of Wheat (*Triticum* species) Kodinar, Gujarat, India. Journal of Environmental Science, Toxicology and Food Technology. 2014; 8: 84- 89.
- 158. Lotschert W and Kohm, HJ. Characteristics of tree bark as an indicator in high emissions areas. Oecologia. 1977; 27: 47-64.