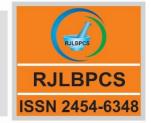
www.rjlbpcs.com



Life Science Informatics Publications

Research Journal of Life Sciences, Bioinformatics, Pharmaceutical and Chemical Sciences

Journal Home page http://www.rjlbpcs.com/



Original Review Article

DOI: 10.26479/2019.0502.05

GREEN BIOSYNTHESIS OF METALLIC NANOPARTICLE FOR MEDICAL DIAGNOSTIC

Swati Sharma, Ragini Gothalwal*

Department of Biotechnology and Bioinformatics Centre, Barkatullah University Bhopal, MP, India.

ABSTRACT: Metal nanoparticles are garnering considerable attention owing to their high potential for use in various applications in biological, biomedical, and biosensing. Nanoparticle have unique Physical and Chemical Properties due to their high surface area and nanosized. The synthesis of metallic nanoparticle has the potential to deliver new source of novel metal that are stable, nontoxic, cost effective, environmental friendly and synthesized using green channel approach. By using microorganisms such as Bacteria, Fungi, Algae/Cyanobacteria and plant, for the production of nanoparticle is an economical, energy efficient and healthier work for human health and environment leading to lesser waste and make useful product, for example, cyanobacterial nanoparticle has been utilized in many medical areas such as hepatocellular carcinoma, nanomaterial-based biosensor, gene therapy, anti-bacterial agent and drug delivery system etc. The review emphasizes the strategies for biosynthesis of metallic nanoparticles using microorganism and plant which have potential to developed novel anti cancerous, antibacterial and nontoxic product.

KEYWORDS: Nanoparticles, Cyanobacteria, Green Synthesis, Drug delivery.

Corresponding Author: Dr. Ragini Gothalwal* Ph.D.

Department of Biotechnology and Bioinformatics Centre, Barkatullah University Bhopal, MP, India. Email Address: raginigothalwal@yahoo.com

1.INTRODUCTION

Metallic Nanoparticles research is currently of intense significance due to a wide variety of potential application in biomedical, drug design and delivery and Medical diagnostic. Nanoparticle are of great scientific interest as they are effectively a bridge between bulk material and atomic or molecular structure. However, nanomedicine present considerable challenges for preclinical application. Nanoparticle are wide class of material that include particulate substance which have

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications one dimension less than 100 nm at least [1]. Depending on the overall shape these materials can be 0D,1D,2D, or 3D [2]. The nanoparticle can be employed for drug delivery [3], chemical and biological sensing [4] [5] and other related application [6]. Biological synthesis of nanoparticles is a one-step bio reduction method and less energy are used for synthesis of ecofriendly nanoparticles. [7]. Nanoparticles are gaining reputation as multifaceted material exhibiting novel or advanced characteristics compound to larger particles. [8]. Smaller size nanoparticles display higher surface to volume ratio a feature vital for catalytic reactivity, thermal conductivity, antimicrobial activity chemical steadiness and non -linear optical performance. [9] because of such characteristics nanoparticles playing significant role in medical diagnostics, antisense and gene therapy application and tissue engineering [10]. In order to provide a more environmental sound synthesis of nanoparticles various biological routes are considered including the use of plant extract [11], enzyme [12], bacteria [13], fungi [14] and algae [15, 8]. Mechanism and protocol for the synthesis of various inorganic metal nanoparticles and nanomaterial have been developed for a wide range of application including biosensor and chemical sensors, bioimaging, catalysis, optics, electronics, drug delivery and energy [16][17]. For example various nanomaterials have been tested as specially controlled carries in drug delivery system for drug transport to the cellular target [18] and use to convert solar energy directly into stream for sanitation and water purification. Several microorganisms such as bacteria, algae, yeast, and fungi, are capable of reducing metal ions through metallo-regulatory mechanism upon exposure [19]. Formation of metal nanoparticles from heavy metal ions occurs through the reduction of the metal ions resulting in the formation of insoluble complexes. This mechanism is employed for the biosynthesis of diverse metal nanoparticles using microorganism engineered to express heavy metal binding protein and /or peptide [20]. The composition of a specific nanoparticles can be very complex, depending on interaction, it has had with other chemicals or particles and its lifetime. The chemical process taking place on surface of nanoparticles are also very complicated and remain largely unknown.

Biosynthesis of Nanoparticles

The principal parameter of nanoparticles is their shape, size, surface characteristics and inner structure. Nanoparticle can be encountered as aerosol [solid or liquid in air] suspension [solid in liquid] or as emulsion [liquid in liquid]. In the presence of certain chemicals properties of nanoparticles may be modified. Biological method for the synthesis of silver nanoparticles employing microorganism [21,22] and plant [23]. Microbial synthesis of nanoparticles does not encounter variation, although regular maintenance of culture and sterile condition for nanoparticles synthesis is required [23].

Bacteria/Fungi Mediated Nanoparticles

The biosynthesis of nanoparticle depends on the cell [24]. Acinetobacter has been shown to exhibit biocompatibility with its own metal nanoparticles however exposure to the corresponding metal,

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications salt decrease their cell count [25]. Fusarium oxysporum has been reported to produce a wide variety of metal nanoparticles including Ag-Au, Ag, cds, cdSe, Co3O4, Sio2, Tio2, ZrO2, BuTio3 and Fe3O4 approximately size 50 to 200 nm [26, 27]. Extra cellular synthesis of nanoparticles occurs outside the bacterial cell such as spherical, disk, cuboidal, hexagonal, triangular etc. have been synthesized using cell culture supernatant or aqueous cell free extract [28]. The intracellular silver nanoparticles depend on the culture medium used to grow the cell, [24]. Individually encapsulated recombinant E. coli cells were able to synthesize homogeneous nanoparticles through concentrated treated metal precursors [16]. Lactobacillus strain has been described to mediated biosynthesis of silver and titanium nanoparticles [29] and gold, silver, and titanium nanoparticle. Vibacterium casei and Aeromonas SH10 silver nanoparticle were synthesized by biomass accumulation as well as Bacillus subtilis, Bacillus licheniformis, Corynebacterium and Pseudomonas studzeri reduced AgNo3 yielding silver nanoparticle produce under atmospheric condition and in size range is 10 and 5 nm, 200 nm respectively [30]. The bacterial strain is used for the synthesis of silver nanoparticles which is isolate from the soil which are identified by Pneumoniae [31]. The synthesis of circular and triangular crystalline silver nanoparticles by solar irradiation of cell free extract of Bacillus amyloliquefaciens and silver nitrate [32], staphylococcus aureus produced biogenic silver nanoparticles against a range of resistant bacteria [33]. The green synthesis of silver nanoparticles from Bacillus sp. Were collected from the periplasmic region of the bacterial cell [34]. Geobacter ferrireducens was also produce gold nanoparticles intracellularly in periplasmic space [35]. In the biological synthetic process for metal nanoparticle soluble sulfate act as the source of sulfur. The formation mechanism of metal nanoparticle by biological transformation mechanism of Rhodobacter spheroids is explained by soluble sulfate which enter into immobilized beads via diffusion and later is carried to the interior membrane of Rhodobacter. Cell facilitated by sulfate permease then the sulfate is reduced to sulfite by ATP sulfurylase and phosphodenamine. Phosphosulfate reductase and next sulfide react with O acetyl serine to synthesize cystine which are solubilize with metal and synthesized metallic nanoparticle. [36].

Algae mediated nanoparticles

Spirulina plantensis synthesized nanoparticles of silver, gold, and gold core, silver shell [Au core-Ag shell]. [37]. An extract of the unicellular green alga Chlorella vulgaris synthesized single crystalline Ag nanoparticles at room temperature extract of the brown alga Chlorella monliformis were shown for the synthesis of silver nanoparticles with size range of 50-100 nm spherical and small particles at low temperature [38,39]. Filamentous cyanobacterium strain i.e. Anabena, Calothrix and Leptolyngbya have the capability to synthesize Au, Ag, Pd and Pt nanoparticles, the microorganism will use for these metallic nanoparticle's formation is Calothrix, Leptolyngbya and Anabena spi. [40]. The mechanism of gold nanoparticles synthesis by cyanobacteria is describe by the interaction of cyanobacteria with aqueous gold chloride initially promoted the precipitation of

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications nanoparticles of amorphous gold sulfide at the cell wall and finally deposited metallic gold in the form of octahedral platelets near cell surface and in solution. This process is based on enzymatic reduction of the metal ions leading to their aggregation and the formation of nanoparticles. [41]. Nanoparticles production using Plectonema boryanum of spherical silver nanoparticles of up to 200 nm is solution [42]. The production of ZnO nanoparticles by the Anabena-flas aquae photosynthetic microorganism [40]. Silver nanoparticles using the leaf extract of Bacillus monosperna were synthesized and observed that at a concentration of 1% leaf extract [43]. Cds nanoparticles were synthesized using C. Phycoerythrin as the capping agent [44]. Silver nanoparticle was synthesized by cyanobacterial and green algae strain in BG 11 medium [45]. Filamentous cyanobacteria strain i.e. Anabena, Calothrix and Leptolyngbya have the capability to form metallic nanoparticles of well controlled size such as silver, gold, platinum, palladium. The algal cell of Spirulina platensis was chosen because it also possessed pharmaceutical and nutraceutical properties for the silver nanoparticles synthesis [37].

Plant Mediated Nanoparticles

Synthesis of gold and silver nanoparticles using dried powder of Catharanthus camphora leaf which are used to check antiplasmodial activities [46]. The extract of the Microbilis jalapa flower work as reducing agent and gold nanoparticles with ecofriendly method [47]. By ecofriendly synthesis of metallic nanoparticles of Piper nigrum leaves and Ag Nps have effective drug in Cancer medicine to cure various oncology and dreadful disease. A green synthesis of silver nanoparticles using the leaves of Artemisia nilagirica plant extract [48]. An ecofriendly method synthesis of gold nanoparticles by using Rose plant is described by Noruzi.et.al., [2011] [49]. Catharanthus -raseus and Cilitora ternate diverse group of flowers are used for the metallic nanoparticle's synthesis with desired size and shape. Nyctanthues arbortristis flower of gold nanoparticles extract are synthesized by green chemical method [50].

Microorganism	and	Types of	Metal	Reference
Plant derived Metallic		microorganism		
Nanoparticles S. No.				
		Bacteria		
1.		Enterobacter aureus	Ag	[51]
2.		Bacillus subtilis	Ag	[52]
3.		Bacillus subtilis	Au	[53]
4.		Cupriavidus	Au	[54]
		metallidurans		
5.		Rhodopseudomans	Au	[55,56]
		capsulate		

Sharma & Gothalwa		5 1	Science Informatics Publications
6.	Bacillus	Te	[57]
7	selenitireducens		[70]
7.	Serratia sps	Cu, Cuo	[58]
8	P. aeruginosa	Ag, Co, Fe, Li, Ni,	[59]
		Pd, Pt, Rh, Ru	
9.	Escherichia coli	Au, Cu, Cuo, Cds	[60], [61]
10.	Sulfate reducing Bacteria	Fe ₃ S ₄ , Fe ₃ O ₄	[62], [63]
11.	Cobalt resistant bacteria	C03O4	[64]
12.	Recombinant E. coli	Au Ag Fe Te Cds	[65], [66], [67], [68]
		Br, Cd, Zn	[69]
13.	Enterobacter	Au, Ag	[70]
13.	aerogens	114, 115	[, 0]
14.	Geobacter	Ag	[71]
1	sufurredicen	1.8	[, 1]
15.	Gluconobacter	Ag	[72]
15.	roseus	116	[,2]
16.	Idiomarina sps	Ag	[73]
17.	Klebsiella	Ag	[74]
17.	pneumoniae	115	[יין
18.	Morganella sps	Ag	[75]
10.	Proteus Microbilis	Ag	[76]
19. 20.	Pseudomonas	Ag	[59]
20.	aeruginosa	Ag	[39]
21.	Yersinia enterocolitis	۸a	[78]
21. 22.	Bacillus	Ag	
22.	thuringenesis	Ag	[79]
23.	Ũ	۸a	[00]
	Rhodococcus sps.	Ag	[80]
24.	Holococcus salifodi	Ag ngi	[81]
25.	Fu Trichoderma viride	-	[00]
		Ag	[82]
26. 27	Coriolus versicolor	Ag	[83]
27.	Volvariella volvacea	Ag	[84]
28.	Penicillium strain	Ag	[85]
29.	Verticillium sps.	Au	[86]
(© 2019 Life Science Informa	atics Publication All rig	hts reserved

	ife Science Informatics Publications
Ag	[87]
Ag	[88]
Ag	[89]
Ag	[90]
	Algae
Au	[91]
Au	[92]
Au, Ag, Pd	[93]
Fe, Mn	[42]
	Plant
Au, Ag	[94]
In_2O_3	[95]
Ag	[96]
Ag	[97]
Ag	[60]
Ag	[98]
Au	[99]
Ag	[100]
Ag	[101]
Ag	[102]
Ag	[103]
Ag	[104]
Ag	[105]
Ag	[49]
Ag	[61]
Ag	[47]
Ag	[49]
	Ag Ag Ag Ag Ag Ag Au Au Au Au Au Au Au Au Ag Ag Ag Ag Ag Ag Ag Ag Ag Ag Ag Ag Ag

Sharma & Gothalwal	RJLBPCS 2019	www.rjlbpcs.com	Life Science Informatics Publications
55.	Iresina herbst	ii Ag	[108]
56.	Melia azedarad	ch Ag	[109]
57.	Tinospora cordif	folia Ag	[110]
58.	Trigonella foeni	um- Au	[111]
	graecum		
59.	Withania somnif	fera Ag	[112]
60.	Mentha Piperi	ta Ag, Au	[44]

Application

Metal nanoparticles have widely and commonly been used in biological, biomedical, and biosensing application. In biological system a large variety of organism from organic/inorganic composites with ordered structure by the use of biopolymers such as protein and microbial cells. Several different approaches for the synthesis of metal nanoparticles based on the different system such as whole live cell versus cell extract, wild type versus recombinant microorganism strain and bulk phase versus microdroplet can be examined such novel metal nanoparticles that have not yet been synthesized can potentially serve as new nanoparticles for exciting industrial application. There are also various ecofriendly nanoparticles product available in commercial market with high efficiency such as Water purifier, bone and teeth cement, facial cream and home-made product [113]. Silver, silica and platinum nanoparticles have various application in personal care and cosmetics and biologically synthesized nanoparticles are used as biological ingredient in various product such as sunscreen, antiaging cream, toothpaste, mouthwash, hair care product and perfume [114]. In medical field tentative evidence support a clearance risk of urinary tract infection, when alloy catheter is used silver nanoparticles synthesized by bacteria exhibit excellent larvicides potency against the dengue vectors, Ades aegypti [115] and Hemophysalis bispinosa [116] indicating their application as a patent insecticide as well as nanomedicine is a field of research with tremendous prospect for the improvement of the diagnosis and treatment of human disease. Dispersed nanoparticles are usually employed in Nano biomedicine as fluorescent biological labels [117], drug and gene delivery agent [118] and in application such as bio detection of pathogens [119], tissue engineering tumor destruction via heating ,MRI contrast enhancement and phagokinetic studies [120], delivering the drug precisely and safely to their target sites at the right time to have a controlled release and achieve the maximum therapeutic effect is a key issue in design and development of novel drug delivery system. Nanoparticles such as silver [121,122], zinc oxide [123,124] and polyethyleneimines nanoparticles [125] are incorporated into dental adhesive [126] or dental composite [127]. The mechanism of action of these antibacterial nanoparticles may vary and include a generation of ROS [128] active transport metabolism of sugar [129] disruption of cell envelope disturbance in membrane electron transport [122]. The application of the biosensing field are known

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications to be ecofriendly and less toxic. It has become a novel and very promising material for nanoelectronics, nanocomposites, opto-electronic devise, electrochemical super capacitor devise, fabricated field effect transistors, drug delivery system, solar cells, memory devices and constructed ultra-sensitive chemical sensors such as PH sensors, gas sensors and biosensors [130,131]. An ultrasensitive electrochemical immunosensor based on nano gold particles [Au-GN-HRP] is used as the label for the immunosensors. An electrochemical immunosensor for the sensitive detection of carbohydrate antigen was fabricated based on ionic liquid functionalized graphene and Cd+2 nanoparticles Tio2 drug biosensors using graphene and Na ion film modified GEC is fabricated for the detection codeine displaying [132].

1.	Aquaspirillum	Au	Biosensors	[133][69]
	magnetatactium		Drug delivery	[93]
			Bioremediation	
2.	Cupriavidus	Ag	Biosensors	[133] [53]
			Drug delivery	[134] [91]
			Bioremediation	[135]
			Antibacterial	
			agents	
3.	Fusarium	Au, Ag	Microscopy,	[136]
	oxysporum		catalysts	
4.	Fusarium	Ag2S	Biofilm	[137]
	oxysporum	BaTiO3	Imaging	[138]
			microelectric	
			device	
5.	Plant stercullia	Cu, CuO	Biosensors	[139]
			Antibacterial	[140]
			agent	[141]
			Cancer therapy	
			Bioremediation	
6.	Clostridium	Co	Bioremediation	[142]
	aceticum		Surface	
			modification	
7.	Pseudomonas	Fe	Cleaning of	[143]
	aeruginosa		contaminated	
			land and water	

Sharma & Gothalwal RJLBPCS 2019		www.rjlbpcs.com	m Life Science	Informatics Publications
8.	Fusarium	Cds	Biosensors	[144,145]
	oxysporum		Drug delivery	[146]
			Electronic	[147]
			devices	
			Biomarker, cell	[148]
			labeling agent	
9.	Pseudomonas	Li	Catalysts	[143]
	aeruginosa			
10.	Plant	Ni	Catalysts	[144]
			Bioremediation	[143]
11.	Bacterial Sps.	Pd	Catalysts	[145]
			Bioremediation	[146]
12.	Plant	Pt	Biosensors	[147]
12.	Fiani	Γl	Antibacterial	
				[148]
			agent Cancer therapy	
			Bioremediation	
13.	Bacillus subtilis	Se	Biosensors	[149]
13.	Bacillus subillis	36	BIOSEIISOIS	[149]
14.	Fusarium	SiO2	Biomedical	[147]
14.		3102		[150]
15	oxysporum Bacillus and	Та	therapy	[151]
15.	Bacillus sps.	Te	Antibacterial	[151]
			agent	

2. CONCLUSION

This review provide a glimpse to some simpler nanoparticles which are being currently modified for their potential application are so beneficial to increase the production which are zero energy consuming and low cost .The synthesis of metallic nanoparticles using biological entities has the potential to deliver , new source of novel material that are stable , nontoxic , cost effective , environment friendly and synthesized using green approach .An increasing awareness towards green chemistry and use green route for the synthesis of metal nanoparticles lead a desire to develop environment friendly technique . Benefit of synthesis of metallic nanoparticles using plant and microorganism is that it is an economical, energy efficient, cost effective provides healthier work place and communities protecting human health and environment leading to lesser waste and safer product. It seems that several important technical challenges must be overcome before this green

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications bio-based method will be a successful and competitive alternative for industrial synthesis of nanoparticles.

ACKNOWLEDGEMENT

Author is thankful to the Head of the Department Biotechnology, Barkatullah university Bhopal for providing the facilities and support during the present work.

REFERENCES

- Laurent. S., Forge. D., Port. M., Roch. A., Robic. C., Vander Elst. L., Muller. R.N. Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. Chem. Rev.2010; 2574–2574.
- Tiwari. J.N., Tiwari. R.N., Kim. K.S. Zero-dimensional, one dimensional, two- dimensional and three-dimensional nanostructured materials for advanced electrochemical energy devices. Prog. Mater Sci. 57, 2012; 724–803.
- 3. Lee. J.E., Lee. N., Kim. T., Kim. J., Hyeon. T. Multifunctional mesoporous silica nanocomposite nanoparticles for theragnostic applications. Acc. Chem. Res. 2011;44, 893–902.
- 4. Barrak. H., Saied. T., Chevallier. P., Laroche. G., M'nif. A., Hamzaoui. A.H., []. Synthesis, characterization, and functionalization of ZnO nanoparticles by N- [trimethoxysilylpropyl] ethylenediamine triacetic acid [TMSEDTA]: investigation of the interactions between phloroglucinol and ZnO@TMSEDTA. Arab. J. Chem. 2016.
- Ullah.H., Khan. I., Yamani. Z.H., Qurashi. A. Sono chemical driven ultrafast facile synthesis of SnO2 nanoparticles: growth mechanism structural electrical and hydrogen gas sensing properties. Ultrason. Sonochem. 2017; 34, 484–490.
- 6. Shaalan. M., Saleh. M., El-Mahd. M., El-Matbouli. M. Recent progress in applications of nanoparticles in fish medicine: a review. Nanomed. Nanotechnol. Biol. Med.2016; 12, 701–710.
- Singaravelu, G., Arokiamary, J.S., Kumar, V.G. and Govindaraju, K.A novel extracellular synthesis of mondisperse gold nanoparticles using marine alga, Sargassum wightti Greville. Colloids Surfaces Biointerfaces, 2007;57[1]: 97-101.
- Rajeshkumar, S., Malarkodi, C., Paulkumar, K., Vanaja, M., Gnanajobitha, G., Annadurai, G. Algae mediated green fabrication of silver nanoparticles and examination of its antifungal activity against clinical pathogens, Int. J. Metals. 2014;1–8.
- 9. Geetha, S., Sathakkathulzariya, J., Aarthi, R. and Blessie, H. Green synthesis of gold nanoparticle using marine cyanobacteria Gloeocapsa sp and the antitumour potential. Journal of Chemical and Pharmaceutical Sciences. 2014; 4: 172- 174.
- Kubik. T., Bogunia-Kubik. K., Sugisaka. M. Nanotechnology on duty in medical applications, Curr. Pharm. Biotechnol. 2005, 6 17–33.
- Gilaki. M. Biosynthesis of silver nanoparticles using plant extracts, J. Biol. Sci. 2010; 10 ,465–467.

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications

- Schneidewind. H., Schuler. T. K., Weber. K., Cialla. D., Diegel M., Mattheis. R., Berger. A., Moller. R., Popp. J. The morphology of silver nanoparticles prepared by enzyme-induced reduction, Beilstein J. Nanotechnol. 2012; 3 ,404–414.
- 13. Saifuddin. N., Wong. C.W., Nur Yasumira. A.A. Rapid biosynthesis of silver nanoparticles using culture supernatants of bacteria with microwave irradiation, E-J. Chem. 2009; 6 ,61–70.
- 14. Balaji.D.S., Basavaraja. S., Bedre. M.D., Prabhakar. B.K., Venkataraman. A. Biosynthesis of silver nanoparticles by fungus Trichoderma reesei, Insci. J. 2011; 1,65–79.
- Jena. J., Pradhan. N., Nayak. Dash. R.R., Sukla. B.P., Panda. L.B., Mishra. P.K., B.K. Microalga Scenedesmus sp.: a potential low-cost green machine for silver nanoparticle synthesis, J. Microbiol. Biotechnol. 2014; 24 522–533.
- Lee. K.G., Hong. J., Wang. K.W., Heo. N.S., Kim. D.H., Lee. S.Y., Lee. S.J., Park. T.J.In vitro biosynthesis of metal nanoparticles in microdroplets. ACS Nano. 2012; 6:6998–7008.
- Yang. M.H., Choi. B.G., Park. T.J., Heo. N.S., Hong. W.H., Lee. S.Y. Site specific immobilization of gold binding polypeptide on gold nanoparticle-coated graphene sheet for biosensor application. Nanoscale. 2011; 3:2950–2956.
- Wilczewska. A.Z., Niemirowicz. K., Markiewicz. K.H., Car. H. Nanoparticles as drug delivery systems. Pharmacol Rep. 2012; 64:1020–1037.
- 19. Kang.S.H., Bozhilov.K.N., Myung. N.V., Mulchandani. A. Microbial synthesis of CdS nanocrystals in genetically engineered E. coli. Angew Chem Int Ed. 2008, 120:5186–5189.
- Park. T.J., Lee. S.J., Kim. D.K., Heo. N.S., Park. J.Y., Lee. S.Y. Development of label-free optical diagnosis for sensitive detection of influenza virus with genetically engineered fusion protein. Talanta. 2012; 89:246–252
- Gaidhani. S., Singh. R., Singh. D., Patel. U., Shevade. K., Yeshvekar. R., Chopade. B.A. Biofilm disruption activity of silver nanoparticles synthesized by Acinetobacter calcoaceticus PUCM. 2013; 108:324–327.
- 22. Singh. R., Wagh. P., Wadhwani.S., Gaidhani. S., Kumbhar. A., Bellare. J., Chopade. B.A. Synthesis, optimization, and characterization of silver nanoparticles from Acinetobacter calcoaceticus and their enhanced antibacterial activity when combined with antibiotics. Int J Nanomedicine. 2013; 8:4277–4290.
- 23. Salunkhe. G.R., Ghosh. S., Santoshkumar. R.J., Khade.S., Vashisth. P., Kale. T., Chopade. S., Pruthi. V., Kundu. G., Bellare. J.R., Chopade. B.A. Rapid efficient synthesis and characterization of silver, gold and bimetallic nanoparticles from the medicinal plant Plumbago zeylanica and their application in biofilm control. Int J Nanomedicine. 2014; 9:2635–2653.
- Srivastava. P., Bragança. J, Ramanan. S.R, Kowshik. M. Synthesis of silver nanoparticles using haloarchaeal isolate Halococcus salifodinae BK3. Extremophiles. 2013; 17:821–831.
- 25. Wadhwani. S.A., Shedbalkar. U.U., Singh. R., Karve.M. S., Chopade. B.A. Novel polyhedral

- Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications gold nanoparticles: green synthesis, optimization and characterization by environmental isolate of Acinetobacter sp. SW30. World J Microbiol Biotechnol. 2014; 30:2723–2731.
- Jha. A.K., Prasad. K., Kulkarni. A.R. Synthesis of TiO2 nanoparticles using microorganism. Colloid Surf B. 2009; 71:226–229.
- Uddin. I., Adyanthaya. S., Syed. A., Selvaraj. K., Ahmad. A., Poddar. P. Structure and microbial synthesis of sub-10 nm Bi2O3 nanocrystals. J Nanosci Nanotechnol. 2008;8:3909–3913.
- Oves. M., Khan. M.S., Zaidi. A., Ahmed. A.S., Ahmed. F., Ahmad. E., Sherwani. A., Owais. M., Azam. A.Antibacterial and cytotoxic efficacy of extracellular silver nanoparticles biofabricated from chromium reducing novel OS4 strain of Stenotrophomonas maltophilia. PLoS One. 2013; 8, e59140.
- Zhang. M., Zhang. K., Gusseme. De. B., Verstraete. W. The antibacterial and anti-biofouling performance of biogenic silver nanoparticles by Lactobacillus fermentum. Biofouling. 2014; 30:347–357.
- 30. Klaus. T., Joerger. R., Olsson. E., Granqvist. C.G. Silver-based crystalline nanoparticles, microbially fabricated. Proc Natl Acad Sci U S A. 1999; 96:13611–13614.
- Duraisamy. K., Yang. S.L. Synthesis and characterization of bactericidal silver nanoparticles using cultural filtrate of simulated microgravity grown Klebsiella pneumoniae. Enzyme Microb Technol. 2013; 52:151–156.
- 32. Wei, X., Luo, M., Xu, L., Zhang, Y., Lin, X., Kong, P., Liu, H. Production of fibrinolytic enzyme from Bacillus amyloliquefaciens by fermentation of chickpeas, with the evaluation of the anticoagulant and antioxidant properties of chickpeas. J. Agric. Food Chem. 2011; 59, 3957– 3963
- Kim, S.H., Lee, H.S., Ryu, D.S., Choi S.J., Lee, D.S. Antibacterial activity of silver-nanoparticles against Staphylococcus aureus and Escherichia coli. Korean J. Microbiol. Biotechnol. 2011; 39: 77-85
- Vaidyanathan, R., Gopalram, S., Kalishwaralal, K., Deepak, V., Pandian, S.R.K., Gurunathan, S. Enhanced silver nanoparticle synthesis by optimization of nitrate reductase activity. 2010; B 75, 335–341.
- Parikh, R. Y., Singh, S., Prasad, B., Patole, M.S., Sastry, M., Shouche, Y.S. Chem. Bio. Chem. 2008;
 9,1415
- 36. Jiang, L. Du, H. Liu, X., Wang, E. Electrochem. Commun. 2007;9,1165.
- Govindaraju, K., Sabjan, K. B., Ganeshkumar, V., Singaravelu,G. Silver, gold and bimetallic nanoparticles production using single-cell protein [Spirulina platensis] geitler. J.Mater. Sci. 2008, 43, 5115–5122.
- 38. Kannan, R.R., Arumugam, R., Ramya, D., Manivannan, K. and Anantharaman. Green synthesis of silver nanoparticles using marine macroalgae Chaetomorpha linum. Applied Nanoscience,

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life S 2013, 3: 229-233.

- Merin, D.D., Prakash, S. and Bhimba, B.V. Antibacterial screening of silver nanoparticles synthesized by marine microalgae. Asian Pacific Journal of Tropical Medicine. 2010, 3[10]:797-799.
- 40. Rauwel, P., Kuunal, S., Ferdov, S. and Rauwel, E. A review on the green synthesis of silver nanoparticles and their morphologies studied via TEM advances. Materials Science and Engineering. 2015;682749: 1-9.
- 41. Lengke, M. F, Southam, G.Bioaccumulation of gold by sulfatereducing bacteria cultured in the presence of gold[I]-thiosulfate complex. Geochim Cosmochim Ac. 2006; 70:3646–3661.
- 42. Lengke. M.F, Fleet. M.E, Southam. G. Biosynthesis of silver nanoparticles by filamentous cyanobacteria from a silver [I] nitrate complex. Langmuir. 2007; 23:2694–2699.
- 43. Chaturvedi, V., Verma, P.Fabrication of silver nanoparticles from leaf extract of Butea monosperma [Flame of Forest] and their inhibitory effect on bloom-forming cyanobacteria a Bioresources and Bioprocessing, 2015;2:18.
- 44. Mubarak, Ali., D., Thajuddin, N., Jeganathan, K., Gunasekaran, M. Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. Colloid Surf. B. 2011;85, 360–365.
- 45. Patel, V., Berthod, D., Puranik, P, Ganter, M. Screening of Cyanobacteria and microalgae for their ability to synthesize silver nanopartiles with antibacterial activity, Biotechnology Report. 2015;112-119.
- 46. Ponarulselvam, S., Panneerselvam C., Murugan K., Aarthi N., Kalimuthu K., Thangamani S. Synthesis of silver nanoparticles using leaves of Catharanthus roseus Linn. G. Don and their antiplasmodial activities, Asian Pac J Trop Biomed 2012; 2[7]: 574-580.
- Vankar, P.S., Bajpai, D.. Preparation of gold nanoparticles from Mirabilis jalapa flowers. Indian J Biochem Biophys. 2010;47[3]:157-60.
- 48. Vijayakumar, M., Priya, K., Nancy, F.T., Noorlidah, A., Ahmed, A.B.A.Biosynthesis, characterisation and anti-bacterial effect of plant-mediated silver nanoparticles using Artemisia nilagirica, Ind Crops Prod. 2013; 41, 235–240.
- Noruzi, M., Zare, D., Khoshnevisan, K., Davoodi, D. Rapid green synthesis of gold nanoparticles using Rosa hybrida petal extract at room temperature. Spectrachem. Acta Part A. 2011; 79, 1461– 1465.
- 50. Das, R.K., Gogoi, N., Bora, U. Green synthesis of gold nanoparticles using Nyctanthes arbortristis flower extract. Bioprocess Biosys. 2011;34, 615–619.
- Sintubin, L., Windt, W., D., Dick, J., Mast, J. Lactic acid bacteria as reducing and capping agent for the fast and efficient production of silver nanoparticles. Appl. Microbiol.Biotechnol. 2009; 84, 741–749.

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications

- 52. Ahmad A, Jagadale T, Dhas V, Khan S, Patil S, Pasricha R, Ravi V, Ogale S. Fungus-based synthesis of chemically difficult-to synthesize multifunctional nanoparticles of CuAlO2. Adv Mater. 2007; 19:3295–3299.
- 53. Southam. G., Lengke. M.F. Bioaccumulation of gold by sulfatereducing bacteria cultured in the presence of gold[I]-thiosulfate complex. Geochim Cosmochim Ac. 2006; 70:3646–3661.
- Reith F, Etschmann B, Grosse C, Moors H, Benotmane M.A, Monsieurs P, Grass G, Doonan C, Vogt S., Lai B., Martinez-Criado G., George, G.N, Nies, D.H, Mergeay, M., Pring A., Southam G., Brugger, J. Mechanisms of gold biomineralization in the bacteriumCupriavidus metallidurans. Proc Natl Acad Sci USA. 2009 106:17757–17762,
- 55. He. S., Guo. Z., Zhang. Y., Zhang. S., Wang. J., Gu. N. Biosynthesis of gold nanoparticles using the bacteria Rhodopseudomonas capsulata. Mater Lett. 2007; 61:3984–3987.
- He. S., Zhang. Y., Guo. Z., Gu. N. Biological synthesis of gold nanowires using extract of Rhodopseudomonas capsulata. BiotechnolProg. 2008; 24:476–480.
- 57. Baesman. S.M., Bullen. T.D., Dewald. J., Zhang. D., Curran. S., Islam. F.S., Beveridge. T.J., Oremland. R.S. Formation of tellurium nanocrystals during anaerobic growth of bacteria that use Te oxyanions as respiratory electron acceptors. Appl Environ Microbiol. 2007; 73:2135–2143.
- Hasan. S.S, Singh. S., Parikh. R.Y., Dharne. M.S., Patole. M.S, Prasad. B.L.V., Shouche.Y.S. Bacterial synthesis of copper/copper oxide nanoparticles. J Nanosci Nanotechnol. 2008; 8:3191– 3196.
- Srivastava S. K, Constanti, M.Room temperature biogenic synthesis of multiple nanoparticles [Ag, Pd, Fe, Rh, Ni, Ru, Pt, Co, and Li] by Pseudomonas aeruginosa SM1. J Nanopart Res. 2012; 14:831.
- Starodub. M.E., Trevors. J.T.Silver accumulation and resistance in Escherichia coli R1. J Inorg Biochem. 1990; 39:317–325.
- Singh, C., Sharma, V., Naik, P.K., Khandelwal, V., Singh, H.A green biogenic approach for synthesis of gold and silver nanoparticles using zingiber officinale. Dig. J. Nanomat. Bios. 2010; 6, 535–542.
- 62. Watson. J.H.P., Ellwood. D.C., Soper.A.K., Charnock. J. Nanosized strongly- magnetic bacterially-produced iron sulfide materials. J Magn Magn Mater. 1999]; 203:69–72.
- 63. Reguera. G., Pollina. R.B., Nicoll. J.S., Lovley. D.R. Possible nonconductive role of Geobacter sulfurreducens pilus nanowires in biofilm formation. J Bacteriol. 2007; 189:2125–2127.
- 64. Kumar. U., Shete. A., Harle. A.S., Kasyutich. O., Schwarzacher.W., Pundle. A., Poddar. Extracellular bacterial synthesis of protein functionalized ferromagnetic Co3O4 nanocrystals and imaging of self-organization of bacterial cells under stress after exposure to metal ions. Chem Mater. 2008b; 20:1484–1491.
- 65. Kang. S.H., Bozhilov. K.N., Myung. N.V., Mulchandani. A., Chen. W. Microbial synthesis of © 2019 Life Science Informatics Publication All rights reserved Peer review under responsibility of Life Science Informatics Publications 2019 Jan – Feb RJLBPCS 5(1) Page No.76

Sharma & Gothalwal RJLBPCS 2019www.rjlbpcs.comLife Science Informatics PublicationsCdS nanocrystals in genetically engineered E. coli. Angew Chem Int Ed. 2008 120:5186–5189.

- 66. Park. T.J., Lee. S.Y., Heo. N.S., Seo. T.S. In vivo synthesis of diverse metal nanoparticles by recombinant Escherichia coli. Angew Chem Int Ed. 2010; 49:7019–7024.
- 67. Jung. J.H., Park. T.J., Lee. S.Y., Seo. T.S.Homogeneous biogenic paramagneticnanoparticle synthesis based on a microfluidic dropletgenerator. Angew Chem Int Ed. 2012; 51:5634–5637.
- 68. Lee. K.G., Hong. J., Wang. K.W., Heo. N.S., Kim. D.H., Lee. S.Y., Lee. S.J., Park. T.J.In vitro biosynthesis of metal nanoparticles in microdroplets. ACS Nano. 2012; 6:6998–7008
- 69. Seo J.M, Kim E.B, Hyun M.S, Kim B.B, Park T.J. Self-assembly of biogenic gold nanoparticles and their use to enhance drug delivery into cells. Colloid Surf B. 2015; 135:27–34.
- 70. Karthik. C., Radha. K.V. Biosynthesis and characterization of silver nanoparticles using Enterobacter aerogenes: a kinetic approach. Dig J Nanomater Biostruct. 2012; 7:1007–1014.
- Law. N., Ansari. S., Livens. F.R., Renshaw. J.C., Lloyd. J.R. The formation of nano-scale elemental silver particles via enzymatic reduction by Geobacter sulfurreducens. Appl Environ Microbiol. 2008; 4:7090–7093.
- Krishnaraj. R.N., Berchmans. S. In vitro antiplatelet activity of silver nanoparticles synthesized using the microorganism Gluconobacter roseus: an AFM-based study. RSC Adv. 2013; 3:8953– 8959.
- Seshadri. S., Prakash. A., Kowshik. M. Biosynthesis of silver nanoparticles by marine bacterium, Idiomarina sp. Bull Mater Sci. 2012; 35:1201–1205.
- 74. Parikh. R.Y., Ramanathan. R., Coloe. P.J., Bhargava. S.K., Patole. M.S., Shouche. Y.S., Bansal.
 V. Genus-wide physicochemical evidence of extracellular crystalline silver nanoparticles biosynthesis by Morganella spp. PLoS One 6. 2011; e21401.
- 75. Samadi. N., Golkaran. D., Eslamifar. A., Jamalifar. H., Fazeli. M.R., Mohseni. F.A. Intra/extracellular biosynthesis of silver nanoparticles by an autochthonous strain of Proteus mirabilis isolated from photographic waste. J Biomed Nanotechnol. 2009; 5:247–253.
- 76. Pourali. P., Baserisalehi.M., Afsharnezhad. S., Behravan. J., Alavi.H., Hosseini. A. Biological synthesis of silver and gold nanoparticles by bacteria in different temperatures [37 °C and 50 °C]. J Pure Appl Microbiol. 2012; 6:757–763.
- 77. Banu,A.N., Balasubramanian, C., Moorthi, P.V. Biosynthesis of silver nanoparticles using Bacillus thuringiensis against dengue vector, Aedes aegypti [Diptera: Culicidae]. Parasitol. 2014; Res 113:311–316.
- Otari, S.V., Patil, R.M., Nadaf, N.H., Ghosh, S.J., Pawar, S.H. Green synthesis of silver nanoparticles by microorganism using organic pollutant: its antimicrobial and catalytic application. Environ Sci Pollut Res. 2014; 21:1503–1513.
- 79. Srivastava, P., Bragança, J, Ramanan, S.R, Kowshik, M.Synthesis of silver nanoparticles using haloarchaeal isolate Halococcus salifodinae BK3. Extremophiles. 2013; 17:821–831.

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications
80. Amanulla, M. F., Balaji, K., Girilal, M., Yadav, R. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: A study against gram-positive and gram-negative bacteria. Nanomed. Nanotechnol. Biol. Med. 2010; 6, 103–109.

- 81. Rashmi, S., Preeti, V. Biomimetic synthesis and characterization of protein capped silver nanoparticles. Biores. Technol. 2009; 100, 501–504.
- Daizy, P. Biosynthesis of Au, Ag and Au–Ag nanoparticles using edible mushroom extract. Spectrochim. Acta A. 2009;73, 374–381.
- Maliszewska, I., Szewczyk, K., Waszak, K. Biological synthesis of silver nanoparticles. J. Phys. Conf. Series. 2009; 146,1–6.
- 84. Priyabrata, M., Ahmad, A., Deendayal, M., Satyajyoti, S. Fungus mediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: a novel biological approach to nanoparticlesynthesis. Nano Lett. 2001; 1, 515–519
- Vigneshwaran, N., Arati, A. K., Varadarajan, P.V., Rajan, P.N. Biomimetics of silver nanoparticles by white rot fungus, Phaenerochaete chrysosporium. Colloids Surface B Biointerface, 2006;53, 55–59.
- 86. Bhainsa, K. C., D'Souza, S. F. Extracellular biosynthesis of silver nanoparticles using the fungus Aspergillus fumigatus. Colloids Surfac B Biointerface 2006, 47, 160–164
- Vigneshwaran, N., Ashtaputre, N. M., Varadarajan, P. V., Nachane, R. P. Biological synthesis of silver nanoparticles using the fungus Aspergillus flavus. Mater. Lett. 2007; 61, 1413–1418.
- 88. Ingle, A., Rai, M., Gade, A., Bawaskar, M. Fusarium solani, A novel biological agent for the extracellular synthesis of silver nanoparticles. J. Nanopart. Res. 2009; 11, 2079–2085.
- Konishi Y, Ohno K, Saitoh N, Nomura T, Nagamine S, Hishida H, Takahashi Y, Uruga T.Bioreductive deposition of platinum nanoparticles on the bacterium Shewanella algae. J Biotechnol. 2007; 128: 648–653.
- 90. Suresh, A.K, Pelletier, D.A, Wang, W., Broich ., M.L, Moon J.W, Gu B, Allison D.P, Joy D.C, Phelps T.J, Doktycz M.J. Biofabrication of discrete spherical gold nanoparticles using the metalreducing bacterium Shewanella oneidensis. Acta Biomater. 2011; 7:2148–2152.
- 91. Keren, N., Aurora, R., Pakrasi, H.B. Critical roles of bacterioterritins in iron storage and proliferation of cyanobacteria. Plant Physiol. 2004; 135: 1666–1673.
- 92. Tripathy, A., Ashok, M. R., Chandrasekaran, N., Prathna, T. C. Process variables in biomimetic synthesis of silver nanoparticles by aqueous extract of Azadirachta indica [Neem] leaves. J. Nanopart. Res. 2010;12, 237-246.
- 93. Maensiri, S., Laokul, P., Klinkaewnarong, J., Phokha, S., Promarak, V., Seraphin, S. Indium oxide [In2O3] nanoparticles using Aloe vera plant extract: synthesis and optical properties. J. Optoelectron. Adv. Mater. 2008;10, 161–165.
- 94. Suriyakalaa, U., Antony, J.J., Suganya, S., Siva, D., Sukirtha, R., Kamalakkannan, S., Pichiah,
 © 2019 Life Science Informatics Publication All rights reserved
 Peer review under responsibility of Life Science Informatics Publications
 2019 Jan Feb RJLBPCS 5(1) Page No.78

- Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications P.B.T., Achiraman, S. Hepatocurative activity of biosynthesized silver nanoparticles fabricated using Andrographis paniculata. Colloid Surf. B. 2013; 102, 189–194.
- Niraimathi, K.L., Sudha, V., Lavanya, R., Brindha, P. Biosynthesis of silver nanoparticles using Alternanthera sessilis [Linn.] extract and their antimicrobial, antioxidant activities.Colloid Surf. B. 2012; 88, 34–39.
- Jain, D., Daima, H.K., Kachhwaha, S., Kothari, S.L.Synthesis of plant-mediated silver nanoparticles using papaya fruit extract and evaluation of their anti-microbial activities. Dig. J. Nanomat. Bios. 2009; 4, 557–563.
- 97. Daisy, P., Saipriya, K. Biochemical analysis of Cassia fistula aqueous extract and phytochemically synthesized gold nanoparticles as hypoglycemic treatment for diabetes mellitus. Int. J. Nanomed. 2012; 7, 1189–1202.
- Sathishkumar, M., Sneha, K S.W.WonC.-W.ChoS.KimY.-S.Yun. Cinnamon zeylanicum bark extract and powder mediated green synthesis of nano-crystalline silver particles and its bactericidal activity, Colloids and Surfaces B: Biointerfaces. 2009; 73, 2, 332- 338.
- Satyavani, K., Gurudeeban, S., Ramanathan, T., Balasubramanian, T. Biomedical potential of silver nanoparticles synthesized from calli cells of Citrullus colocynthis [L.] Schrad. J. Nanobiotechnol. 2011; 9, 43–51.
- 100. Kaviya, S., Santhanalakshmi, J., Viswanathan, B., Muthumary, J., Srinivasan, K. Biosynthesis of silver nanoparticles using Citrus sinensis peel extract and its antibacterial activity. Spectrochim. Acta A. 2011; 79, 594–598.
- 101. Singh, S., Saikia, J.P., Buragohain, A.K.A novel 'green' synthesis of colloidal silver nanoparticles [SNP] using Dillenia indica fruit extract. Colloid Surf. B. 2013; 102, 83–85.
- 102. Ghosh, S., Patil, S., Ahire, M., Kitture, R., Kale, S., Pardesi, K., Cameotra, S.S., Bellare, J., Dhavale, D.D., Jabgunde, A., Chopad, B.A.Synthesis of silver nanoparticles using Dioscorea bulbifera tuber extract and evaluation of its synergistic potential in combination with antimicrobial agents. Int. J. Nanomed. 2012; 7, 483–496.
- 103. Zahir, A.A., Rahuman, A.A. Evaluation of different extracts and synthesized silver nanoparticles from leaves of Euphorbia prostrate against the plant Haemaphysalis bispinosa and Hippobosca maculate. Vet. Parasitol. 2012; 187, 511–520.
- 104. Cruz, D., Fale', P.L., Mourato, A., Vaz, P.D., Serralheiro, M.L., Lino, A.R.L.Preparation and physicochemical characterization of Ag nanoparticles biosynthesized by Lippia citriodora [Lemon Verbena]. Colloid Surf. B. 2010; 81, 67–73.
- 105. Dipankar, C., Murugan, S. The green synthesis, characterization and evaluation of the biological activities of silver nanoparticles synthesized from Iresine herbstii leaf aqueous extracts. Colloid Surf. B. 2012; 98, 112–119.
- 106. Sukirtha, R., Priyanka, K.M., Antony, J.J., Kamalakkannan, S., Thangam, R., Gunasekaran, P.,
 © 2019 Life Science Informatics Publication All rights reserved
 Peer review under responsibility of Life Science Informatics Publications
 2019 Jan Feb RJLBPCS 5(1) Page No.79

- Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications Krishnan, M., Achiraman, S. Cytotoxic effect of Green synthesized silver nanoparticles using Melia azedarach against in vitro HeLa cell lines and lymphoma mice model. Process Biochem. 2012; 47, 273–279.
- 107. Jayaseelan, C., Rahuman, A.A., Rajakumar, G., Kirthi, A.V., Santhoshkumar, T., Marimuthu, S.Synthesis of pediculocidal and larvicidal silver nanoparticles by leaf extract from heart leafmoon seed plant Tinospora cordifolia Miers. Parasitol. Res. 2011; 109,185–194.
- 108. Aromal, S.A., Philip, D.Green synthesis of gold nanoparticles using Trigonella foenumgraecum and its size dependent catalytic activity. Spectrochim. Acta A. 2012; 97, 1–5.
- 109. Nagati, V.B., Alwala, J., Koyyati, R., Donda, M.R., Banala, R., Padigya, P.R.M. Green Synthesis of plant-mediated silver nanoparticles using Withania somnifera leaf extract and evaluation of their anti-microbial activity. Asian Pac. J. Trop. Biomed. 2012; 2, 1–5.
- Kouvaris, P., Delimitis, A., Zaspalis, V., Papadopoulos, D., Tsipas, S.AMichailidis, N. Green synthesis and characterization of silver nanoparticles produced using Arbutus unedo leaf extract. Mater. Lett. 2012; 76, 18–20
- 111. Kumar, V., Yadav, S.K. Plant-mediated synthesis of silver and gold nanoparticles and their applications. J. Chem. Technol. Biot. 2009;84, 151–157.
- 112. Debabov V.G, Voeikova T.A, Shebanova A.S, Shaitan K.V, Emel'yanova L.K, Novikova L.M, Kirpichnikov M.P. Bacterial synthesis of silver sulfide nanoparticles. Nanotechnol Russ. 2013; 8:269–276.
- Karthik L, Kumar G, Kirthi AV, Rahuman AA, Rao KVB. Streptomyces sp. LK3 mediated synthesis of silver nanoparticles and its biomedical application. Bioprocess Biosyst Eng. 2014; 37:261–267.Colloids and Surfaces B: Biointerfaces. 111, 1.2013; 680-687.
- 114. Bruchez, M, Moronne, M, Gin P, Weiss, S, Alivisatos, A.P. Semiconductor nanocrystals as fluorescent biological labels. Science. 2016; 281.
- 115. Sahu, P.K., Iyer, P.S., Barage, S.H., Sonawane, K.D., Chopade, B.A. Characterization of the algC gene expression pattern in the multidrug resistant Acinetobacter baumannii AIIMS 7 and correlation with biofilm development on abiotic surface. Sci World J. 2014;10. 1155
- 116. Dey, R. S., Hajra, S., Sahu, R. K., Raj, R. C. and Panigrahi, M.K.A rapid room temperature chemical route for the synthesis ofgraphene: metal-mediated reduction of graphene oxide. Chem. Commun. 2012; 48, 1787–1789.
- 117. Liu, S., Wang, J., Zeng, J., Ou, J., Li, Z., Liu, X. and Yang, S. 'Green' electrochemical synthesis of Pt/graphene sheetnanocomposite film and its electrocatalytic property. J. PowerSources. 2010; 195, 4628–4633.
- 118. Blecher, K., Nasir, A., Friedman, A. The growing role of nanotechnology in combating infectious disease. Virulence. 2011; 2 [5], 395–401.
- 119. Li, Q.M.S., Lyon, D.Y., Brunet, L., Liga, M.V., Li, D., Alvarez, P.J.Antimicrobial nanomaterials
 © 2019 Life Science Informatics Publication All rights reserved
 Peer review under responsibility of Life Science Informatics Publications

2019 Jan – Feb RJLBPCS 5(1) Page No.80

- Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications for water disinfection and microbial control: potential applications and implications. Water Res. 2008; 42 [18], 4591–4602
- Gu, H., Fan, D., Gao, J.Effect of ZnCl2 on plaque growth and biofilm vitality. Arch. Oral Biol. 2012; 57 [4], 369–375.
- 121. Xie, Y., He, Y., Irwin, P.L., Jin, T., Shi, X. Antibacterial activity and mechanism of action of zinc oxide nanoparticles against Campylobacter jejuni. Appl. Environ. Microbiol.2011; 77 [7], 2325–2331.
- 122. Shvero, D.K., Davidi, M.P., Weiss, E.I., Srerer, N., Beyth, N. Antibacterial effect of polyethyleneimine nanoparticles incorporated in provisional cements against Streptococcus mutans. J. Biomed. Mater. Res. 2010; 94 [2], 367–371.
- 123. Chen, C., Weir, M.D., Cheng, L. Antibacterial activity and ion release of bonding agent containing amorphous calcium phosphate nanoparticles. Dent. Mater. 2014; 30 [8], 891–901.
- 124. Kasraei, S., Sami, L., Hendi, S., Alikhani, M.Y., Rezaei, L., Khamverdi, Z. Antibacterial properties of composite resins incorporating silver and zinc oxide nanoparticles on Streptococcus mutans and Lactobacillus. Restor. Dent. Endod. 2014; 39 [2], 109–114.
- 125. Sobjerg, L.S., Gauthier, D., Lindhardt, A.T., Bunge, M., Finster, K., Meyer, R.L., Skrydstrup, T. Bio-supported palladium nanoparticles as a catalyst for Suzuki–Miyaura and Mizoroki–Heck reaction. Green Chem. 2009; 11:2041–2046
- 126. Kawabata, N., Nishiguchi, M. Antibacterial activity of soluble pyridiniumtype polymers. Appl. Environ. 1998;54, 2532–2535.
- 127. Gurunathan, S., Han, J. W., Dayem, A. A., Eppakayala, V. andKim, J.-H. Oxidative stressmediated antibacterial activity ofgraphene oxide and reduced graphene oxide in Pseudomonasaeruginosa. Int. J. Nanomedicine. 2012; 7, 5901–5914.
- 128. Gurunathan, S., Han, J. W., Eppakayala, V. and Kim, J.-H. Green synthesis of graphene and its cytotoxic effects in humanbreast cancer cells. Int. J. Nanomed. 2013; 8, 1015–1027.
- 129. Gupta, S. S., Sreeprasad, T. S., Maliyekkal, S. M., Das, S. K. and Pradeep, T. Graphene from sugar andits application in waterpurification. ACS Appl.Mater. Interfaces. 2012;4, 4156–4163.
- Mann, S., Frankel, R.B., Blakemore, R.P. Structure, morphology, and crystal growth of bacterial magnetite. Nature. 1984; 310:405–407.
- Kashefi K, Tor J.M., Nevin K.P., Lovley, D.R. Reductive precipitation of gold by dissimilatory Fe [III]-reducing bacteria and archaea. Appl Environ Microbiol.2001; 67:3275–3279,
- 132. Fayaz, A.M., Balaji, K., Girilal, M., Yadav, R., Kalaichelvan, P.T., Venketesan, R. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against Gram-positive and Gramnegative bacteria. Nanomed-Nanotechno. 2010;16:103–109
- 133. Senapati, S., Ahmad, A., Khan, M.I., Sastry, M., Kumar, R. Extracellular biosynthesis of bimetallic Au-Ag alloy nanoparticles. Small. 2005; 1:517–520.

Sharma & Gothalwal RJLBPCS 2019 www.rjlbpcs.com Life Science Informatics Publications 134. Sastry, M, Ahmad, A, Khan, M.I, Kumar, R. Microbial nanoparticle production. In: NiemeyerCM, Mirkin CA [ed]. Nanobiotechnology. 1st edn. Wiley-VCH Verlag GmbH & Co.,Weinheim. 2004; 126–135.

- 135. Jha, A.K, Prasad, K. Ferroelectric BaTiO3 nanoparticles: Biosynthesis and characterization. Colloid Surf B. 2010; 75:330–334.
- 136. Padil, V.V.T., Cernik, M., [2013], Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. Int J Nanomedicine 8:889–898.
- Iravani, S. Green synthesis of metal nanoparticles using plants. Green Chem. 2011; 13:2638– 2650.
- 138. Guo, H., Luo, S., Chen, L., Xiao, X., Xi, Q., Wei, W., Zeng, G., Liu, C., Wana, Y., Chen, J., He, Y. Bioremediation of heavy metals by growing Appl Microbiol Biotechnol. 2010.
- Korbekandi, H., Iravani, S., Abbasi, S. Production of nanoparticles using organisms. Crit Rev Biotechnol. 2009; 29:279–306.
- 140. Nies DH, Microbial heavy-metal resistance. App Microbiol Biotechnol ;[1999],51:730-750.
- 141. Yong P, Rowsen NA, Farr JPG, Harris IR, Macaskie LE, Bioreduction and biocrystallization of palladium by Desulfovibriodesulfuricans. Biotechnol Bioeng[2002] 80:369–379.
- 142. Kowshik, M., Deshmukh, N., Vogel, W., Urban, J., Kulkarni, S.K., Paknikar, K.M. Microbial synthesis of semiconductor CdS nanoparticles, their characterization, and their use in the fabrication of an ideal diode. Biotechnol Bioeng. 2002a; 78:583–588
- 143. Kowshik, M., Vogel, W., Urban, J., Kulkarni, S.K., Paknikar, K.M. Microbial synthesis of semiconductor PbS nanocrystallites. Adv Mater. 2002b; 14:815–818.
- 144. Mohanpuria, P., Rana, N.K., Yadav, S.K. Biosynthesis of nanoparticles: technological concepts and future applications. J Nanopart Res. 2008; 10:507–517.
- 145. Mandal, D., Bolander, M.E., Mukhopadhyay, D., Sarkar, G., Mukherjee, P. The use of microorganisms for the formation of metal nanoparticles and their application. Appl Microbiol Biotechnol. 2006; 69:485–492.
- 146. Li, X., Xu, H., Chen, Z.H., Chen, G.Biosynthesis of nanoparticles by microorganisms and their applications. J Nanomater. 2011;1–16.
- Bhattacharya, R., Mukherjee, P. Biological properties of Bnaked metal nanoparticles. Adv Drug Deliv Rev. 2008; 60:1289–1306.
- 148. Wang, T., Yang, L., Zhang, B., Liua, J. Extracellular biosynthesis and transformation of selenium nanoparticles and application in H2O2 biosensor. Colloid Surf B; 2010 80:94–102.
- 149. Jha, A.K., Prasad, K., Kulkarni, A.R.Synthesis of TiO2 nanoparticles using microorganism. Colloid Surf B. 2009; 71:226–229.